



INTERNATIONAL APPLICATION PUBLISHED UNDER THE PATENT COOPERATION TREATY (PCT)

<p>(51) International Patent Classification ⁷ : G01N 27/26, 27/447</p>	<p>A1</p>	<p>(11) International Publication Number: WO 00/36407</p> <p>(43) International Publication Date: 22 June 2000 (22.06.00)</p>
<p>(21) International Application Number: PCT/US99/29195</p> <p>(22) International Filing Date: 9 December 1999 (09.12.99)</p> <p>(30) Priority Data: 09/212,622 16 December 1998 (16.12.98) US</p> <p>(71) Applicant: CURAGEN CORPORATION [US/US]; 555 Long Wharf Drive, New Haven, CT 06511 (US).</p> <p>(72) Inventors: BADER, Joel, S.; Apartment 4E, 85 South Main Street, Branford, CT 06405 (US). ROTHBERG, Jonathan, M.; 45B Cocheco Avenue, Branford, CT 06405 (US). DEEM, Michael, W.; 24 Highland Avenue, #14, Cambridge, MA 02139 (US). MULHERN, Gregory, T.; Apartment #1, 45 Harrison Avenue, Branford, CT 06405 (US). WENT, Gregory, T.; 25 Scotland Avenue, Madison, CT 06443 (US). SIMPSON, John; Apartment B4, 23 Woodland Road, Madison, CT 06443 (US). HENCK, Steven; 11 Maple Vale Drive, Woodbridge, CT 06525 (US).</p> <p>(74) Agents: ANTLE, Adriane, M. et al.; Pennie & Edmonds LLP, 1155 Avenue of the Americas, New York, NY 10036 (US).</p>		<p>(81) Designated States: AE, AL, AM, AT, AU, AZ, BA, BB, BG, BR, BY, CA, CH, CN, CR, CU, CZ, DE, DK, EE, ES, FI, GB, GD, GE, GH, GM, HR, HU, ID, IL, IN, IS, JP, KE, KG, KP, KR, KZ, LC, LK, LR, LS, LT, LU, LV, MA, MD, MG, MK, MN, MW, MX, NO, NZ, PL, PT, RO, RU, SD, SE, SG, SI, SK, SL, TJ, TM, TR, TT, UA, UG, UZ, VN, YU, ZA, ZW, ARIPO patent (GH, GM, KE, LS, MW, SD, SL, SZ, TZ, UG, ZW), Eurasian patent (AM, AZ, BY, KG, KZ, MD, RU, TJ, TM), European patent (AT, BE, CH, CY, DE, DK, ES, FI, FR, GB, GR, IE, IT, LU, MC, NL, PT, SE), OAPI patent (BF, BJ, CF, CG, CI, CM, GA, GN, GW, ML, MR, NE, SN, TD, TG).</p> <p>Published With international search report.</p>
<p>(54) Title: SEPARATION OF CHARGED PARTICLES BY A SPATIALLY AND TEMPORALLY VARYING ELECTRIC FIELD</p>		
<p>(57) Abstract</p> <p>This invention relates to a method and device for separating particles according to their diffusivities in a separation medium by means of a spatially and temporally varying electric potential. The method takes advantage of the transport of charged particles subject to an electric potential that is cycled between an off-state (in which the potential is flat) and one or more on-states, in which the potential is preferably spatially periodic with a plurality of eccentrically shaped stationary wells. The potential wells are at a constant spatial positions in the on-state. Differences in liquid-phase diffusivities lead to charged particle separation. A separation medium fills physically defined separation lanes (15) in the device. Electrodes deposited substantially transverse to the lanes create the required potential. Advantageously, injection ports (16) allow sample loading, and special gating electrodes focus the sample prior to separation.</p>		

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SEPARATION OF CHARGED PARTICLES BY A SPATIALLY AND TEMPORALLY VARYING ELECTRIC FIELD

This is a continuation-in-part of U.S. patent application Ser. No. 08/623,346, filed
5 March 27, 1996, which is incorporated by reference herein in its entirety.

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This invention was made with United States Government support under award
number 70NANB5H1036 awarded by the National Institute of Standards and Technology.
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1. FIELD OF THE INVENTION

This invention relates to a method and apparatus for the separation of charged
particles in a medium according to the differences of the diffusivities of the particles in the
20 medium by use of a spatially and temporally varying electric potential. Particularly, the
invention relates to a method and apparatus for separation of charged biopolymers in a
liquid medium, and more particularly to a method and apparatus for the separation of
single-stranded or double-stranded DNA fragments for DNA sequencing and for general
fragment length determination.

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This invention further relates to a method and apparatus for using electrode and
optical design patterns, and other methods of device construction, to enhance the utility of
this diffusivity-based technique to cause the motion or separation of charged species, such
as DNA.

30

2. BACKGROUND OF THE INVENTION

Separations of charged particles, in particular physical mixtures of chemical species,
are important analytical operations. Relevant chemical species include non-biological
charged species, such as synthetic polymers, and biological charged species, such as DNA,
35 RNA, or proteins (A.J. Kostichka et al., 1992, *Bio/Technology* 10:78). Separations of
mixtures of DNA fragments are particularly important.

For example, the Human Genome Project demonstrates the need for powerful DNA
fragment separation methods and apparatus. This project is an ambitious, international
effort to improve human genetic maps, to sequence fully the genomes of humans and

several model organisms by 2006, and to develop computational tools for storing and accessing the burgeoning information. This project requires a technological infrastructure capable of supplying high-quality sequence information in a rapid and cost-effective manner.

5 To sequence fully the human genome, which has approximately 3×10^9 base pairs, by the year 2006 requires roughly 100 times beyond the total, current worldwide DNA sequencing capacity (M.V. Olson, 1993, *Proc. Natl. Acad. Sci. USA* 90:4338). Existing DNA sequencing methods, for example, mass spectrometry (T.D. Wood et al., 1995, *Proc. Natl. Acad. Sci. USA* 92:11451), sequencing by hybridization (R. Drmanac et al., 1993, ¹⁰ *Science* 260:1649), chromatography (C.G. Huber et al., 1993, *Nucl. Acids Res.* 21:1061), acoustophoresis (J.S. Heyman, U.S. Patent No. 5,192,450), and electrophoresis, are generally inadequate to meet this sequencing goal.

The above methods have various drawbacks. Mass spectrometry requires an expensive mass spectrometer. Because of this cost, it is unlikely that this method will have ¹⁵ widespread applicability. Sequencing by hybridization is still relatively new and untested. Liquid chromatography is capable of performing rapid separation of double-stranded DNA fragments, but is limited by poor resolution. The single-base resolution necessary for sequencing has only been demonstrated for fragments smaller than 150 base pairs. In ²⁰ acoustophoresis, acoustic waves push fragments through a liquid medium. This method is limited by the similarity in the acoustic properties of DNA fragments of similar lengths, preventing effective separation.

Electrophoresis remains the most common method by far for DNA sequencing. All conventional electrophoretic methods are generally similar (F. Sanger et al., 1977, *Proc.* ²⁵ *Natl. Acad. Sci. USA* 74:5463; L.M. Smith, 1993, *Science* 262:530). A DNA sample is generally first amplified, that is the DNA chains are made to replicate, usually by the polymerase chain reaction ("PCR"). Next, from the amplified sample, chain terminating DNA polymerase reactions (first described by Sanger et al.) produce nested sets of DNA ³⁰ fragments labeled with one of four unique fluorescent dyes conjugated with one of the four chain terminating bases (either ddATP, ddCTP, ddGTP, or ddTTP). In a related method, the chains are cleaved by chemical means to produce a similar set of labeled fragments (M. Maxam et al., 1977, *Proc. Natl. Acad. Sci. USA* 74:560). These fragments are then separated according to their molecular size by a variety of electrophoretic techniques, and ³⁵ the unique dye labeling each chain terminating base is detected by its fluorescence. The DNA base sequence is reconstructed from the detected pattern of chain fragments.

The accuracy required in DNA fragment size determination depends on the application. For example, DNA sequencing reactions produce a mixture, called a "ladder," of fragments with lengths separated by single bases and require exact length determination.

Other applications produce greater differences between the fragment lengths, and methods that provide rapid sizing, but not necessarily exact length information, are valuable. Typical of such applications are the generation of patterns of restriction fragment length polymorphism ("RFLP"), genotyping, linkage analysis, microsatellite analysis and other
5 fragment analysis application.

In an electrophoretic separation, the DNA molecules are separated according to their rates of migration in an electric field. The electric driving force is proportional to the net charge of the molecule. For a uniformly charged biopolymer such as DNA, the driving force is proportional to the number of base pairs in the DNA fragment. Since in a material
10 obeying Stokes' Law, such as a liquid, the friction coefficient is also proportional to the number of base pairs, the DNA fragments have electrophoretic drift velocities that are nearly identical and independent of fragment length. This means electrophoretic separation of DNA fragments is difficult in liquids or other media obeying Stokes' Law.

Therefore, instead of liquid media, cross-linked gels and uncross-linked polymer
15 solutions are universally used in electrophoretic DNA separations. In these media, DNA does not obey Stokes' law, since the electrophoretic drift velocity decreases with increasing length or molecular weight. Thus, electrophoretic separation of biopolymers is ordinarily performed in a polymeric gel, such as agarose or polyacrylamide, in which separation of
20 biopolymers with similar electric charge densities, such as DNA or RNA, depends on molecular weight. The non-Stokes' law dependence of the friction coefficient on the fragment size in a gel permits electrophoretic separation of DNA fragments of different lengths. Biopolymer fragments, therefore, exit the device in size order from small to large.

In a prevalent configuration, the electrophoretic gel is disposed as a thin sheet
25 between two flat, parallel, rectangular glass plates. An electric field is established along the long axis of the rectangular configuration, and molecular migration is arranged to occur simultaneously in several paths, or "lanes," parallel to the electric field. To ensure high separation resolution, it is advantageous that gel throughout a migration lane be as uniform
30 as possible (or homogeneous like a liquid) and for the lanes to be sufficiently separated to be clearly distinguishable.

It has proven difficult to make, or "to cast," uniform gels with uniform transport properties. One major problem is uneven gel shrinkage due to cross-linking during gel polymerization. The problems in casting a uniform gel also lead to difficulties in producing
35 a uniform and reproducible loading region, into which sample mixtures are placed prior to separation. It is generally accepted that a separation medium with more reproducible transport properties (*i.e.*, more like a homogeneous liquid) would have great utility.

In addition to high separation resolution, demands for more rapid electrophoresis have created additional problems for gel manipulation. Rapid electrophoresis is desirable

for rapid, high capacity biopolymer analysis. This requires, primarily, stronger electric fields that exert greater forces on migrating molecules in order to move them at greater velocities. However, higher fields, voltages, and velocities lead to increased resistive heating in the gel, and consequently, significant thermal gradients in the gel. Such thermal
5 gradients cause additional gel non-uniformities that further impair separation resolution.

To maintain resolution at higher voltages, ever smaller gel geometries are used so that damaging heat may be more readily conducted away. Thus, electrophoresis has been described in geometries where the parallel glass plates are spaced from 25 to 150 μm apart, instead of the usual spacings which are typically greater than 1000 μm (A.J. Kostichka et
10 al., 1992, *Bio/Technology* 10:78). It has proven even more difficult to cast uniform gels of such thinness and to assure long, parallel, narrow, and closely spaced migration lanes in so thin a sheet.

In turn, to overcome these difficulties in thin gels, physical separation means have
15 been used to keep lanes distinct. These separation means create yet a further set of problems. In one such method for producing physically distinct lanes, arrays of capillary tubes with diameters down to 100 μm have been used (X.C. Huang et al., 1992, *Anal. Chem.* 64:2149). These capillary arrays are difficult to cast with uniform gels and difficult to load with samples of fragments. Easy loading is advantageous to minimize the time and
20 cost of the separation setup, which is often labor-intensive. An alternative is to use a dilute polymer solution instead of a gel in each capillary (P.D. Grossman, U.S. Patent No. 5,374,527). However, single base resolution in such solutions has been limited to DNA chains with fewer than 200 bases and loading the capillaries with samples remains difficult
25 (A.E. Barron et al., 1993, *J. Chromatogr. A* 652:3; A.E. Barron et al., 1994, *Electrophoresis* 15:597; and Y. Kim et al., 1994, *Anal. Chem.* 66:1168). Other alternatives include producing physically distinct lanes by microfabrication of channels in an electrophoretic device (D.J. Harrison et al., 1992, *Anal. Chem.* 64:1926 and D.J. Harrison et al., 1993, *Science* 261:895). Electrodes can be deposited to provide precise control of the
30 electrophoretic field (G.T.A. Kovacs et al., 1990, European Patent 0 376 611 A3 and D.S. Soane et al., U.S. Patent No. 5,126,022). In another alternative to migration through gels, optical microlithography has been used to fabricate a quasi-two-dimensional array of migration obstacles for the electrophoretic separation of DNA (W.D. Volkmuth et al., 1992, *Nature* 358:600).

35 Small lane size coupled with the desirability of separating many samples in many migration lanes at once creates conflicting physical requirements. Simultaneous detection of fragments migrating in multiple lanes requires a spatially compact disposition of the migration lanes in order that all the lanes can be observed at once by a spectrograph of limited aperture. However, loading samples into migration lanes prior to separation

requires physical access to the migration lanes that is easier and more rapid for widely spaced lanes. Conventional, flat-plate techniques have only straight, parallel lanes and cannot accommodate these divergent requirements.

Such problems with prior gel-based electrophoretic separation methods have
5 motivated a search for new separation methods. A non-electrophoretic method for separation of particles that are electrically polarizable, but not charged, is based on differences in diffusivities in liquid of the particles. Only mega-base size DNA fragments have sufficient polarizability to be separated by this method (A. Ajdari et al., 1991, *Proc. Natl. Acad. Sci. USA* 88:4468; J. Rousselet et al., 1994, *Nature* 370:446; and J.F. Chauwin
10 et al, 1994, *Europhys. Lett.* 27:421). This method uses an electric field that is periodic but asymmetric in space, substantially transverse to the direction of separation, and cycles temporally from on to off. When the asymmetric field is turned on, it attracts and traps polarizable particles into a series of spatially periodic attractive regions according to the
15 known laws of electrostatics. When the potential is turned off, however, the particles are free to diffuse. Since smaller particles diffuse more rapidly, the cycling electric field causes a size separation of polarizable particles.

The polarization-based device is suited for separating particles on the order of the size of viruses, and may also be able to effect the separation of mega-base fragments of
20 DNA, such as entire chromosomes (J. Rousselet et al., 1994, *Nature* 370:446). This particle size limitation is due to the requirement that the particles to be separated have polarizabilities sufficiently large to be attracted by fields that can be realistically created in a liquid. Since the attractive force varies as the square of the electric field, high voltages are
25 needed. Separation of DNA fragments of a few 100's of bases in length, the sizes commonly produced by sequencing reactions or by RFLPs, is out of reach of this or similar polarization-based devices due to practical limits on electric field strength and voltages.

All the foregoing technical problems have hindered creation of a machine for rapid, concurrent analysis of large numbers of biopolymer samples at low cost and with minimal
30 human intervention. The need for such a machine is widely felt in many areas of biology such as, for example, biological research, the Human Genome Project, the biotechnology industry, and clinical diagnosis.

Citation of references hereinabove shall not be construed as an admission that such reference is prior art to the present invention.

35

3. SUMMARY OF THE INVENTION

It is one object of the method and apparatus of this invention to provide convenient and efficient separation of charged particles which overcomes the problems in the prior art. The particles to be separated may be charged either positively or negatively. In particular,

the charged particles separated can be biopolymer fragments, such as single-stranded or double-stranded DNA.

It is another object of the method and apparatus of this invention that separation is accomplished with a spatially and temporally varying electric potential. The potential has a plurality of eccentrically-shaped potential wells that trap the charged particles when the potential is relatively strong. Separation occurs as particles diffuse differentially from well to well, based on differences in diffusivity in the separation medium, when the potential is sufficiently weak (or off). It is an advantage that there is no overall electric potential difference along the line of particle separation.

The invention provides designs for electrode patterns that enhance the use of the technique of the invention for separation and motion of charged particles. Some of these designs enable superior formation of initial packets of concentrated sample prior to initiation of separation, thereby enhancing separation resolution. Other designs provide a substantially uniform surface material within the separation channel, with minimum contact to electrodes, enabling a choice of material that is highly compatible with the charged particles (such as DNA). Still further designs allow for refocusing of widely distributed samples following separations to enhance detectability. In one such design, the charged particles are continuously refocused to a centerline of the separation channel.

Alternative modes of operation of the invention include temperature ramping and voltage ramping. In one embodiment, the temperature of the device is changed during the course of operation. This is advantageous for separating groups of particles that have widely varying diffusion coefficients. In another embodiment, the potential of the device is varied during operation. In this embodiment, separation is effected as a result of the different response to the voltage variation by differently sized fragments.

It is possible to combine various aspects of the invention. One method of the invention for separating charged particles in a separation medium is first to focus the particles, then to divide the particles into a plurality of species, and finally to refocus each of the species. Embodiments of the invention include use of the spatially and temporally varying electric fields to achieve the initial focusing, to achieve the division into individual species, to achieve the refocusing, or to achieve any combination of these. The electrode designs described above allow selective use of the spatially and temporally varying electric fields to achieve these different embodiments of the invention.

It is an advantage of the method and apparatus that charged particles interact with an electric potential. This is a stronger interaction, for readily obtainable electric fields, than the weaker interaction between polarizable particles and an inhomogeneous electric field, which depends on the square of the electric field strength, the degree of spatial inhomogeneity of the field, and the polarizability of the particles.

It is an advantage of the method and apparatus of this invention that the form of the electric potential of this invention separates charged particles solely on the basis of their diffusivity in the separation medium. Thereby, this invention can separate DNA fragments in liquid or Stokes' Law medium, which is not possible for conventional electrophoresis.

- 5 Further, more rapid DNA separation is possible, as liquid diffusivities are larger than gel diffusivities. However, the method and apparatus of the invention are not limited to liquid separation media.

It is an advantage of the method and apparatus that a liquid medium may be used instead of a gel-based medium. When using a liquid medium, the method and apparatus of
10 the invention is free of many of the limitations of gel-based media, including, for example, difficulty in loading gels in small geometries, gel non-uniformities due to shrinkage, electro-endosmosis, and inhomogeneous gel casting. Because a liquid separation medium is substantially uniform, more reproducible separation is possible and non-uniformities in the
15 separation medium are minimized. Further, the apparatus is quickly reusable. Samples may be removed by applying a uniform high voltage. Alternatively, the liquid separation medium may be quickly flushed, and the apparatus then washed with cleaning solution and refilled with fresh liquid medium.

It is an advantage that the apparatus of the invention can be of small scale suitable
20 for microfabrication. The small scale results in high throughput. A small scale results in efficient heat transfer, reducing separation medium non-uniformities due to local heating. Further, the smaller the apparatus the more rapid the separation. Moreover, the apparatus is suitable for low cost microfabrication. Multiple lanes can be fabricated on a single one
25 centimeter square substrate. A separation module according to this invention can be integrated with sample preparation and fragment detection apparatus. A loading zone can be fabricated on the separation module including electrodes generating a loading potential for localizing loaded samples into a compact volume prior to separation.

It is an advantage of the method and apparatus of the invention that operating
30 parameters can be adjusted to the sizes of the molecules to be separated and the separation resolution required. Thus more rapid separation can be obtained if all the molecules are short or if only approximate sizes (5%-10% accuracy) are required.

It is an advantage of the method and apparatus of the invention that they can provide
35 the superiorities over the conventional electrophoresis systems listed in Table 1.

Comparison	This Invention	Conventional Electrophoresis
Materials	Inexpensive to build and operate; relatively non-toxic separation medium (either aqueous buffer or denaturing organics may be used)	Expensive to operate; separation medium generally uses hazardous polymers and polymerizing agents
Set-Up	Easy to load: separation medium may be liquid	Difficult to load: requires mixing, pouring, and polymerizing a gel
Loading	Focusing by loading zone electrodes yields a narrow initial sample distribution	Diffusion in loading region can yield a broad initial sample distribution and broad bands
Speed	250 bases/hour/lane for a 0.3 μ m feature size (current limit of microfabrication technology)	100-200 bases/hour on a commercial device
Resolution	Bands are reproducible due to a homogeneous separation medium and migration lanes that are physically etched	Bands can lack reproducibility due to inhomogeneities in gel separation medium and curving of separation lanes in gel slab
Multiplexing	100 lanes fit easily on a chip 1 cm square	Multiple lanes in small geometries can be difficult to resolve
Safety	Low voltage operation	High voltage operation
Clean-Up	Easy to flush device with new separation medium and re-use	Requires disposal of toxic gel and cleaning of supporting plates

Table 1: Superiority Over Conventional Electrophoretic Systems.

These objects and advantages are achieved by an invention that separates charged particles, in particular charged chemical species, in a separation medium according to differences in the diffusivities of the particles in the medium by use of spatially and temporally variable electric potential. The spatial variations of the electric potential create along the line of separation a plurality of potential wells that attract and trap the charged particles. The potential wells are eccentrically-shaped, with potential minima disposed off-center with respect to the well. In one embodiment, the wells are generally saw-tooth shaped, having one side that is generally steeper than the other side. The potential wells can be disposed in various spatial configurations, with a configuration periodic along the line of separation being preferred. Temporally, the electric potential cycles between at least two states, with two states being preferred. In at least one of the states, the "on-state," the

particles are attracted to and trapped in the potential wells. In at least another of the states, the "off-state," the particles are substantially free to diffuse according to their diffusivities in the separation medium. In a preferred embodiment, the on-state has a duration sufficient to localize each particle in some potential well, and the duration of the off-state is optimized to provide the most rapid separation possible. Thereby, as the potential cycles between the temporal states, the particles diffuse from potential well to potential well in a predictable manner according to the diffusivities and are, thereby, separated according to their diffusivities.

In an important application of the method, the particles are charged biopolymers. In particular, separation of DNA in a medium, such as a liquid, is important, for example, in DNA sequencing and in observing restriction fragment length polymorphism patterns ("RFLP"), genotyping, linkage analysis, microsatellite analysis and other DNA analysis applications. The method is applicable to DNA separation because single-stranded and double-stranded DNA molecules are charged species with liquid-phase diffusivities depending substantially only on their fragment length.

The method and apparatus of this invention are effective with a wide variety of electric fields having spatial variations creating a plurality of potential wells eccentrically placed with respect to their adjacent maximums along the line of separation, and having temporal variations between at least one state trapping the charged particles in the wells and at least one state permitting substantially free diffusion in the medium (which is preferably liquid medium). The potential wells may have a steep and a less steep side, may have an eccentric shape with a narrow minimum for closely confining the trapped particles, or may have a general asymmetric and eccentric shape. The potential wells may be disposed periodically or with varying distance along the line of separation. A preferred potential is of a saw-tooth form along the line of separation. Temporally, the potential may vary between more than two states or may vary continuously. The temporal variation may be constant during a separation or may change during a separation. A preferred potential varies only between an on-state and an off-state.

As the subsequent disclosure makes apparent, the parameters defining the spatial and temporal variation of the electric potential can be selected in view of the diffusivities and charges of the particles to be separated so that the apparatus can be of any physical size. However, in a preferred embodiment, and especially for the separation of DNA fragments, the apparatus is constructed to achieve the fastest possible separation. In such an embodiment, the device is as small as can be constructed using available microfabrication technology.

An embodiment of the separation apparatus comprises a module containing one or more non-communicating separation lanes for holding the separation medium and along

which the DNA fragments are separated. The module is constructed from two substrates of centimeter ("cm") scale. One substrate is flat, and the other has channels created by, for example, etching grooves or by depositing walls. When the two substrates are joined, the separation lanes are thereby formed.

5 Alternate separation lane geometries are possible. One geometry has straight, parallel lanes. A preferred geometry has lanes spaced widely at a loading zone of the module, in order to permit easy physical access to the lanes for loading, but spaced closely at a detection end, in order to permit simultaneous detection of separated fragments in all the separation lanes. Channel sizes can be less than 1 mm, 500 μm , or 100 μm , and can be
10 as small as 25 μm .

The spatially and temporally varying electric potential is created in a preferred embodiment of the separation module by electrodes that are deposited on whichever of the substrates is flat (that does not have grooves). In a preferred embodiment, electrodes lie
15 substantially transverse to said channels and are disposed to create spatially periodic potential wells, each well having a generally eccentric "sawtooth" shape. In this preferred embodiment a voltage difference is applied to the electrodes for an "on" time, t_{on} , and the electrodes are at the same potential for an "off" time, t_{off} . The potential difference and the "on" time are chosen as sufficient to localize and trap the charged fragments in the potential
20 wells. The "off" time is chosen so that the fragments have a finite probability to diffuse to the next potential well. Cycling the potential causes separation of the charged fragments based on differences in diffusivities. The detailed description (Section 5) makes apparent how to choose the various operational parameters.

The separation medium is chosen to meet several criteria. First, the particles to be
25 separated must be charged in the medium and preferably have a wide range of diffusivities. Second, the medium should both have a high electrical breakdown potential gradient and also not be easily electrolyzed. Preferably, the separation medium is a liquid. Examples of such media appropriate for separating DNA include aqueous liquid media, aqueous buffer
30 solutions, and non-aqueous denaturing liquid media, such as formamide. The invention is not limited to a liquid separation medium. Any media with appropriate electrical properties and in which the particles to be separated are charged and have varying diffusivities can be employed, such as various gels or polymers of various concentrations.

Various enhancements and alternatives in the basic separation module are
35 contemplated by this invention. In an embodiment, liquid separation medium and samples for separation can be loaded through injection ports, which are holes created, for example, by drilling in one of the substrate plates of the separation module. To accommodate such injection ports, the separation lanes may need to be more widely spaced in their vicinity. In addition, an embodiment of the apparatus can include special electrodes to create separate

gating potential wells which serve to localize and trap the samples loaded into the loading ports into a compact initial volume prior to separation.

Control of temperature and temperature gradients in the apparatus is desirable and is preferably achieved with a thermal control module in good thermal contact with one or both
5 substrates. An apparatus of the preferred small size provides especially good thermal control, since the small separation medium channels are necessarily in good thermal contact on all sides with both substrate plates. In an embodiment of the apparatus, the thermal control module comprises bi-directional heat transfer devices, such as Peltier thermo-electric modules, arranged for pumping heat in either direction between the separation
10 module and a heat sink which, for example, exchanges heat with an air or water exchange fluid.

In a preferred embodiment, observation of separated particles is accomplished by optical methods. One possible such optical observation method comprises labeling the
15 particles with unique fluorescent tags, generating a fluorescent signal by laser or other excitation transverse to the separation lanes, and detecting the tag fluorescence with standard spectrometers. A transmission imaging spectrograph may be advantageously used to detect fluorescence simultaneously from multiple separation lanes. The invention is particularly adapted to DNA sequence analysis, in which each DNA molecule is labelled
20 with a different one of four spectrally distinctive fluorescent dyes conjugated to one of the four chain terminating ddNTPs. It is also similarly applicable to applications in which particles to be separated are labeled with multiple dyes.

Numerous modifications that could be made to this apparatus by one skilled in the relevant art are contemplated by this invention. Some of these modifications include the
25 following. The temperature of the media can be varied to enhance diffusivity. Separation media such as polymer solutions or gels can also be used. A variety of materials can be used as the substrate of the separation module and the electrical components, such as the insulators and conductors of the apparatus. Different electrode geometries could be used to
30 obtain electric potentials that function to create potential wells. For example, electrodes may be deposited as a layer contacting the bottom of the separation lane; they may be thicker, extending across the thickness of the separation lane; or they may have an intermediate thickness. Alternately, the potential wells can be created by electrodes external to the separation lanes. A variety of lane geometries are possible, including linear,
35 piece-wise linear, open curvilinear, or closed curvilinear geometries. In a circular geometry, the lanes run around the circumference of a cylinder.

The method and apparatus of this invention has utility in many areas. Biological research laboratories need easy-to-use systems for high-throughput, multiplexed DNA analysis for genome sequencing. Medical laboratories also have growing needs for rapid,

low-cost DNA analysis and sequencing. Separation of other charged particles, for example RNA and proteins, has similar uses in research and diagnostic laboratories.

4. BRIEF DESCRIPTIONS OF THE DRAWINGS

5 These and other objects, features, and advantages of the invention will become apparent to those of skill in the art in view of the accompanying drawings, detailed description, and appended claims, where:

Fig. 1 illustrates a separation device according to the present invention;

10 Fig. 2 illustrates an exploded view of an embodiment of type I of the separation device of Fig. 1;

Fig. 3 illustrates a cross sectional view transverse to the direction of separation of the device of Fig. 2;

15 Figs. 4A-4B illustrates in detail the electrodes of the device of Fig. 2 and the electric potential generated by the electrodes;

Fig. 5 illustrates a cross sectional view along the direction of separation of the device of Fig. 2;

Fig. 6 illustrates the loading zone of the device of Fig. 2;

20 Fig. 7 illustrates an exploded view of an embodiment of type II of the separation device of Fig. 1;

Fig. 8 illustrates a cross sectional view transverse to the direction of separation of the device of Fig. 7;

25 Figs. 9A-9E illustrate in summary form the operation of a method of the present invention;

Fig. 10 illustrates a form of electric potential adaptable for use in the method of Figs. 9A-9E;

Figs. 11A-11D illustrate in detail form the behavior of the particle concentration profile in two adjacent potential wells in the method of Figs. 9A-9E;

30 Figs. 12A-12E illustrate in detail form the behavior of the particle concentration profile in a plurality of adjacent potential wells in the method of Figs. 9A-9E;

Fig. 13 illustrates the behavior of T_{tot} versus t_{off} for the preferred method for the selection of the operating parameters of the method of Figs. 9A-9E;

35 Fig. 14 illustrates the behavior of T_{tot} versus the percentage of separation resolution of DNA molecules when method operating parameters are selected according to the preferred method for the selection of the operating parameters of the method of Figs. 9A-9E;

Fig. 15 illustrates an exemplary photolithography mask for the fabrication of electrodes for the device of Fig. 2;

Figs. 16A-16B illustrate an exemplary photolithography masks for the fabrication of channels for the device of Fig. 2; and

Fig. 17 illustrates an example of a hypothetical separation of DNA molecules according to the method of Figs. 9A-9E.

5 Fig. 18 illustrates the quad-symmetric interdigitated electrode array with center focus gap.

Fig. 19 illustrates an embodiment of the quad-symmetric design in which analyte is continuously driven toward the diagonal centerline of the device.

10 Fig. 20a illustrates the design concept of separate electrode section for use in post-separation sample refocusing.

Fig. 20b illustrates the detection advantage associated with the electrode design shown in Fig. 20a.

15 Fig. 21 illustrates an embodiment of the device in which the separation channel walls are all of similar material, with minimal electrode contact to the analyte.

Fig. 22 illustrates a preferred embodiment incorporating advantages shown in Figures 18 through 20.

5. DETAILED DESCRIPTION OF THE INVENTION

20 Sec. 5.1 describes the structure of an exemplary separation device according to this invention. Sec. 5.2 describes in a summary fashion the operation of a separation method and device. Sec. 5.3 describes in more detail the operation of the separation method and device, and provides a method for the selection of method operating conditions and device
25 Sec. 5.4 describes the important case of the separation of DNA. Finally, Sec. 5.5 describes exemplary methods for microfabrication a separation device according to this invention.

5.1. Description of A Separation Device

30 Fig. 1 illustrates a separation device according to this invention. This separation device and its particular embodiments, device types I and II, are the preferred physical structures for the device. However, charged particle separation according to the method of this invention can be practiced in any other physical structure having one or more separation
35 lanes that hold a separation medium and that are subject to electric potentials according to the method of the invention. For example, it can be practiced in one or more tubes, perhaps of capillary size, with externally imposed electric potentials. Alternatively, the invention can be practiced in an conventional configuration with the separation medium disposed as a slab with the separation lanes running in the slab.

In Fig. 1, separation device 10 includes two substrates, upper substrate 11 and lower substrate 12, which define between them one or more separation lanes, such as lane 15. Lane 15 extends between loading port 16, which, for example, could be a hole drilled in upper substrate 11, and an observation zone indicated generally at 18. In the case where the charged particles are fluorescently tagged, observation can be conveniently effected by illumination of the observation zone 18 by laser beam 19. Fluorescence 20 induced in particles in the separation lanes is observed by a spectrometer. Standard optics together with a CCD detector can detect 0.01 femtomole ("fm") of fluorescent dye-labeled particles in an illuminated area 100 μm on a side. See, e.g., J.W. Simpson *et al.*, "Apparatus and method for the generation, separation, detection and recognition of biopolymer fragments," U.S. Patent Application Serial No. 08/438,231 filed May 9, 1995, which is herein incorporated by reference in its entirety.

The electric potential according to a method of this invention is generated along channel 15 by a pattern of electrodes, to be described hereinbelow, deposited or affixed to one or both substrates. Each electrode is connected to one of electrode pads 13 or 14, through which it is energized by a time varying voltage conducted from external voltage source 17.

Operational parameters of the method can be selected so that the device may be constructed to be of any size. For faster charged particle separation, it is preferable to make the device as small as possible within the constraints of a chosen fabrication or micro-fabrication technology. Small size also permits placing an array of channels on single substrates and thereby achieving multiple simultaneous separations. However, the device should be sufficiently large both so that the charged particles to be separated move independently of each other during separation and also so that at least several hundred particles can be transported through the device together. In the case where the charged particles are charged molecules, this latter condition is easily satisfied by making the smallest dimensions in the device larger than 50 to 500 times the molecular dimensions.

Many materials can be used for substrates, channel walls (in type II devices), and electrodes. One limitation is that any material exposed to a separation lane should be substantially inert to contents of the lane, such as the separation medium used and charged particles to be separated. Either the materials should be inherently inert or should be protected through a passivation layer, for example a silicon oxide layer deposited over electrodes. Further, for ease of detecting the separated particles, it is preferable that the substrate permit the use of fluorescent labels. Thus the substrate should be substantially transparent to light at the excitation and induced fluorescence wavelengths. Amorphous silicon oxide is an example of an acceptable substrate for most fluorescent dyes, such as

those conventionally used to label DNA fragments. More examples of acceptable materials are found in Sec 5.5.

The charged particles to be separated can range from individual molecules of all sizes, to complexes of any number and kind of molecules, and to particles of macroscopic
5 dimensions.

The separation lanes are filled with a separation medium that preferably displays certain properties. A first property is that it dissolves and ionizes the particles to be separated. A second property is that the types of charged particles to be separated have different diffusivities in the medium; the greater the diffusivities overall and the greater the
10 diffusivity difference the more preferable the medium. A third property for rapid separation is that the medium withstands large potential gradients. Therefore, the medium is preferably resistant to electrolysis at the potentials imposed, and the higher the breakdown electric field the better. Finally, it is preferable that the medium have a small dielectric
15 constant and a small ionic strength since the strength of an electric field is reduced by the dielectric shielding due to the medium and by ionic shielding due to contained ions. This latter property is, of course, constrained by the necessity to solvate charged molecules, which generally requires a high dielectric solvent and a finite ionic strength due to the presence of counterions.

20 Separation mediums meeting these conditions for a particular type of charged particle to be separated can be most readily determined by experimentation. For separating biopolymers, such as DNA, suitable separation media are the aqueous solvent or aqueous buffers commonly used in conventional electrophoresis. Alternatively, the medium may also be a denaturing solvent like formamide. Other organic solvents that can solvate
25 molecules in a charged state and permit sufficient potential gradients, for instance DMSO (dimethyl sulfoxide) or alcohol solutions, are also useable. Further, dilute gels or polymer solutions are also possible separation mediums.

The separation device may be operated at any temperature compatible with its
30 construction materials and with the separation medium used. All examples and calculations herein assume operation at ambient temperature, approximately 298 K, unless otherwise stated. Regardless of the operational temperature, it is important that thermal gradients be minimized to keep the separation medium substantially uniform throughout each separation lane. This can be achieved by placing the top or bottom substrate, or both, of the separation
35 device in thermal contact with heat sinks or sources, as appropriate.

Two particular embodiments of the separation device illustrated in Figure 1, called types I and II, are further described in the subsections which follow. In device type I, channels forming the separation lanes, such as channel 15, are etched on one side of one of the substrates 11 or 12. In device type II, the preferred embodiment, separation channels

are formed on one of the substrates by depositing parallel channel walls. In both embodiments, electrodes are deposited on one of the substrates.

5.1.1. Device Type I

5 Fig. 2 is an exploded view of an exemplary embodiment of device type I. Device 10 includes top substrate 11 and bottom substrate 12. One or more separation channels 15 are fabricated in top substrate 11, for example, by etching or micro-machining. Separation occurs along the channels, which, therefore, define separation direction S. The geometry of a channel is preferably approximately semi-circular to rectangular with a width of
10 approximately 50 μm and a height of 10 μm . Smaller heights and widths are preferable because less sample is required for analysis. The width may range to a large distance, comparable to the widths of traditional gel-based electrophoresis devices. The height is preferably sufficiently small that the electric potentials generated are sufficient to localize
15 the particles during the on-condition, a condition most readily determined by experimentation for a given device geometry using particles representative of those to be separated. Each channel extends for substantially the entire length of the substrates, which is typically from 1 to 10 cm, but is chosen according to the design methods of Sec. 5.3. and 5.4. The channels are spaced apart as closely as possible, and are preferably spaced apart no
20 less than a distance approximately equal to their width. Top plate 11 may be fabricated with drilled loading port 16 in Fig. 1, as further described below. The diameter of such a port is selected to permit loading into the lanes of the particles to be separated. The channels may be substantially parallel or, alternately, may converge from a wide spacing in the vicinity of the loading port to a narrow spacing in the observation zone. The two
25 substrates are bonded together by thermal fusing, by the use of adhesives, or by other means of attachment so that the channels fabricated in the top plate are sealed to create closed particle separation lanes. The top substrate has recessions with respect to the bottom substrate to allow the electrode pads to be exposed in order to make electrical connection to
30 external voltage source 17.

Two pluralities of interdigitated electrodes situated facing each other and connected electrode pads 13 and 14, each plurality of electrodes connected to one of the two electrode pads, are deposited on the flat upper surface of bottom substrate 12. Alternatively, electrodes can be deposited on the non-flat surfaces of upper substrate 11. Electrodes
35 and 22 are exemplary of that plurality of electrodes connected to pad 13, and electrodes 21 and 23 are exemplary of that plurality connected to pad 14. These electrodes preferably extend substantially transverse to separation axis S. Less preferably, the electrodes are inclined with respect to the separation axis and separation lanes, as would be the configuration with converging separation lanes. The greater the angle of inclination, the

less efficient and the lower the resolution of the particle separation. The thickness of each of these electrodes is preferably less than approximately 0.1-0.2 μm , and less preferably less than 1 μm , although large sizes will not necessarily interfere with the operation of the device. The width of each of these electrodes, their feature size denoted R' , is preferably less than approximately 1-2 μm . Larger values for R' will not interfere with the operation of the device, but will result in less preferable operation times scaling as R'^2 . Feature sizes of approximately 1 μm are readily achievable with standard micro-fabrication techniques. Preferably, each plurality of electrodes attached to electrode pads 13 and 14 are periodically positioned with a uniform separation of L . For example, electrodes 20 and 22 are separated by distance L , as are electrodes 21 and 23. Preferably, each plurality of electrodes is displaced with respect to one another by a displacement of R . For example, the centers of electrodes 20 and 21 are separated by distance R , as are the centers of electrodes 22 and 23. Preferably R is approximately equal to R' , while L is chosen so that R/L is less than approximately 0.1, although ratios up to 0.5 may be employed. Methods for optimally choosing R , L , and R/L , in terms of feature size R' , achievable in a selected fabrication technology, are described in Sec. 6.2. Alternatively, R , L or R/L can be systematically varied along a separation axis to optimize certain aspects of partial separation according to the model described in Sec. 5.2 and 5.3.

Fig. 3 illustrates a cross sectional view of device 10 along axis 3-3, which is transverse to separation direction S . Top substrate 11 and bottom substrate 12 form three channels, such as channel 15, which are sealed to form separation lanes. Exemplary electrode 20 extends along the bottom of the channels, preferably covered with a passivation layer, if needed.

Fig. 4A schematically illustrates the two pluralities of the electrodes in more detail. Electrodes 20 and 22 of one plurality of electrodes are attached to electrode pad 13, while electrodes 21 and 23 of the other plurality are attached to pad 14. Electrodes of one plurality are separated by distance L . The centers of neighboring electrodes, one of each plurality, are separated by distance R . Each electrode has width R' . Therefore the adjacent edges of electrodes 21 and 22 are separated by distance $L-2R-R'$, and the adjacent edges of electrodes 20 and 21 (or 22 and 23) are separated by $R-R'$. Pads 13 and 14 are charged to potential $-V_0/2$ and $+V_0/2$, respectively. When $V_0 > 0$, the direction of separation of positively charged particles is S . When $V_0 < 0$, S is the direction of separation of negatively charged particles.

Fig. 4B illustrates the approximate, idealized electric potential generated by the two pluralities of electrodes as observed along a separation lane. The potential consists of a series of potential wells, varying from a minimum of $-V_0/2$ in the vicinity of the electrodes attached to pad 13 to a maximum of $+V_0/2$ in the vicinity of the electrodes attached to pad

14. The potential is generally of a saw-tooth shape, periodic in space, with each period or potential well having a uniformly and eccentrically placed minimum. Each period has a relatively shorter and more steeply rising portion 32, between positions 36 and 37 separated by distance R , and a relatively longer and more slowly falling portion between positions 35 and 36 separated by distance $L-R$. The direction of separation, arrow S , is the direction from one minimum to its nearest adjacent maximum. Thus, arrow S is in the direction from minimum 36 towards its nearest adjacent maximum 37. The potential wells are uniform in that these directions for all the wells are all aligned in the same direction, here arrow S . Further, it is clear that the potential wells remain stationary in space in the vicinity of the generating electrodes. In case the pads are charged to opposite potentials, the rising and falling portions are interchanged.

The minimum size for each potential well is generally limited by the preferred aspect that it contain at least several hundred of the particles to be separated. The well should also be large enough to allow the contained particles to move independently without any correlations. In the preferred application in which charged biopolymers are to be separated, this is satisfied if R is larger than approximately $0.1\text{ }\mu\text{m}$.

Fig. 5 illustrates a cross sectional view of device 10 along axis 4-4 of Fig. 2, the direction of separation, in a separation lane, such as lane 15. The separation lane is bounded above by top substrate 11 and below by bottom substrate 12, which are separated by H , the lane height. The lane height is preferably chosen to be $10\text{ }\mu\text{m}$, with larger heights possible subject to the constraint that the electric potential be sufficiently strong to localize the particles during the on-condition. Electrodes, generally at 20, 21, 22, and 23, on bottom plate 12, are substantially transverse to the separation direction S , and are exposed in the separation lane to generate the potential. These electrodes are of height d , preferably less than approximately $0.1\text{-}0.2\text{ }\mu\text{m}$, of width R' , preferably from $1\text{-}2\text{ }\mu\text{m}$, and of separation R , preferably approximately $2R'$, with smaller distances more preferable since they afford more rapid separations. The electrodes are preferably periodically placed with a periodicity distance L , preferably chosen so that R/L is 0.1 or less. Alternatively, R/L is less preferably less than 0.3 ; the device continues to function, albeit less efficiently, up to a limiting ratio of 0.5 (*i.e.* symmetric wells).

The device can be advantageously adapted to have a loading zone for easy loading of particles prior to separation. To allow easy loading with current loading technologies, the loading ports preferably have a diameter of the order of $50\text{-}100\text{ }\mu\text{m}$, the size of micropipettes. Correspondingly, this is a convenient scale for the width and separation of the separation lanes. Alternatively, narrower separation lanes can be widely spaced in a loading zone to accept ports of the preferable size and can converge to a narrower spacing in an observation zone. To achieve optimum separation resolution and speed, it is preferable

that all the particles are attracted into a single potential well prior to separation and that the spacings between sequential potential wells be as previously described.

Fig. 6, an expanded and exploded view of device 10 about loading port 16 of Fig. 1, illustrates a loading zone adapted to meet these properties. Particles to be separated are introduced from outside the device, at position 51, through pipette 50, or similar mechanism, to the interior of separation lane 15, at position 52. Electrodes 54 and 55 underneath port 16 have an increased separation of the order of the diameter of port 16. During or after particle loading, a potential is applied to the electrode pads, and thus to electrodes 54 and 55, for a time sufficient to attract all the particles into the close vicinity of electrode 55, at position 53. A sufficient time can be estimated in a manner similar to the determination of t_{on} described in Sec 5.3. After the particles have been attracted and trapped, particle separation can begin. Alternative electrode configurations may be used in the loading region to achieve a smaller initial distribution of particles to be separated. For example, electrode 56 may be separately held at a potential more attractive than that of electrode 55 to localize all particles between these electrodes prior to beginning separation.

5.1.2. Device Type II

Fig. 7 is an exploded view of an exemplary embodiment of device type II, the preferred embodiment of the separation device. Device 10 includes top substrate 11 and bottom substrate 12. A pattern of electrodes and connected electrode pads similar to that of device type I is deposited on the flat upper surface of bottom substrate 12. Alternatively, electrodes can be deposited on the non-flat surfaces after the channel walls have been fabricated or on the bottom surface of upper substrate 11.

The only difference between the two device types is that in device type II separation lanes are formed by fabricating substantially straight channel walls along direction of separation S on one of the two substrates. Fig. 7 illustrates channel walls 41 and 42 forming separation lane 15 fabricated on the upper surface of the bottom substrate, on top of the electrode pattern previously fabricated. As in device type I, the separation lanes are exposed to the electrodes, which run preferably substantially transverse to the direction of separation S and less preferably have an angle of inclination less than 48°. The geometry of the separation lanes is substantially rectangular with dimensions similar to the lane dimensions of device type I.

The top and bottom substrates are bonded together by thermal fusing, by the use of adhesives, or by other means of attachment so that the channels walls together with the top and bottom substrates form sealed and closed separation lanes. The top substrate has offsets with respect to the bottom substrate to allow the pads to be exposed in order to make an electrical connection with voltage source 17.

Fig. 8 illustrates a cross sectional view of device 10 along axis 8-8 in Fig. 7, which is transverse to separation direction S. Top substrate 11 and bottom substrate 12 bound three channels. Channel 15 is bounded by wall 41 and 42 fabricated on the surface of one of the substrates. Exemplary electrode 20 extends along the bottom of the channels.

5

5.2. Summary Operation of Method

A method of this invention, which is implemented in the devices described in Sec. 5.1, is illustrated in Figs. 9A-E. These figures illustrate the separation of two types of charged particles, a larger particle type indicated by larger rods, as at 91, and a smaller
 10 particle type indicated by smaller rods, as at 92. The electric potential is depicted by curves 90. It assumes a saw-tooth shape for a time t_{on} in Figs. 9A, 9C, and 9E, and is flat for a time t_{off} in Figs. 9B and 9D. In the case where these particles are single-strand DNA molecules of various sizes, the molecules are in reality more likely to be globular in shape.

Fig. 9A represents the beginning of a separation at which time all the particles are
 15 trapped in the left-most potential well. In Fig. 9B, the potential is flat for t_{off} , during which time the particles diffuse equally in both directions along the separation channel. The diffusion is indicated generally at 93. In Fig. 9C, the potential again assumes a saw-tooth shape, and particles that have drifted to the right at least a distance R to the next potential
 20 well are attracted to and trapped in the middle well. However, particles that have diffused less than a distance R are attracted to and trapped in the original left-most potential well. Since smaller particles with larger diffusion constants are more likely to diffuse further than larger particles with smaller diffusion constants, two of the small particles but only one of
 25 the large particles arrive in the middle potential well. In Fig. 9D, the potential is again flat and the particles diffuse equally in both directions from both potential wells at 93 and 94. Finally, in Fig. 9E, when the potential again assumes a saw-tooth shape, one of the small particles has diffused from the middle well far enough to be attracted to and trapped in the right-most well, and two small particles are in the middle well. On the other hand, no large
 30 particles have diffused far enough to be in the right-most well, and only one large particle is in the middle well. It can be seen, therefore, that the particles with the higher diffusion constants will be selectively transported to the right through the device.

The differential forward motion of the particles is due to their diffusion. The potential wells remain spatially stationary, and when on, only serve to attract particles into
 35 their minimum. If the distance R is too great for significant diffusion during the time the potential is off, the particles remain stationary in the device.

In particular, this method can separate DNA molecules because the diffusion constant, D , of DNA depends predictably on the molecular dimensions, and thus on the number of bases, N , in single-stranded or double-stranded fragments (Doi et al., 1986, *The*

Theory of Polymer Dynamics, Clarendon Press, Oxford, p. 300). Experimental measurements of dsDNA and theoretical prediction for ssDNA show that for aqueous solutions:

$$\begin{aligned} D_{dsDNA} &\approx 1.14 \times 10^{-6} N^{-1} cm^2/s \\ D_{ssDNA} &\approx 1.14 \times 10^{-6} N^{-0.59} cm^2/s \end{aligned} \quad (1)$$

See, e.g., Weast, ed., 1987, *Handbook of Chemistry and Physics*, Chemical Rubber Publishing Co., Boca Raton, FL, and Sec. 5.4.

We now consider several of the operating conditions for the device. The separation speed of a potential depends on the its eccentricity, the more eccentric the faster the separation. Eccentricity refers to the location of the potential minimum with respect to the potential well, the closer the potential minimum is to the nearest adjacent maximum the more eccentric is the potential. For example, for a series of saw-tooth potentials with the same period, L , the potential with the smallest R/L ratio operates fastest. Of course, R cannot be substantially smaller than the feature size, R' , available in the chosen fabrication technology, and cannot be so small that the resulting potential gradient exceeds the breakdown field of the separation medium. Also, L is preferably large enough that the potential well can trap at least several hundred independently moving particles.

The voltage, V , applied across the electrode pads should preferably be sufficiently large that t_{on} is as small as reasonably possible compared to t_{off} . However, it should not be so large that substantial electrolysis occurs at the electrodes, that the breakdown field of the separation medium is exceeded, or that resistive heating of the separation medium interferes with separation resolution.

In Sec. 5.3, methods are provided for selecting R , L , R/L , t_{on} , t_{off} , and V based on a model of the separation method in a narrow channel with substantially transverse electrodes. Operating parameters for an actual device should be correctly predicted to within an order of magnitude by this model. If needed, precise operating parameters can be determined from the predicted parameters by routine experimental optimization. For example, in the case of separation of DNA molecules, operation of the device with a DNA standard containing a ladder of fragments of known lengths can be used to optimize the predicted operating parameters.

The method of this invention is adaptable to charged particles of all sizes. The charged particles to be separated can range from individual molecules of all sizes, to complexes of any number and kind of molecules, and to particles of macroscopic dimensions.

5.3. Detailed Operation Of A Method

INTRODUCTION

In this section, the operation of a method of this invention is described in more detail. This description makes use of the following variables:

- 5 L the spatial period of the electric potential;
- R the distance from a potential minimum to the nearest potential maximum (the extent to which R is less than $L/2$ is a measure of the eccentricity of each period of the potential);
- P the temporal period of the electric potential ($P = t_{on} + t_{off}$);
- 10 f the temporal frequency of the electric potential ($f = 1/P$);
- t_{on} the time when the potential is applied, during which the particles are attracted and trapped in the potential wells;
- t_{off} the time when the potential is not applied, during which the particles can freely diffuse;
- 15 Q the charge of a charged particle;
- V_o the applied potential difference;
- T the temperature;
- D the diffusion constant of one type of charged particle to be separated; and
- 20 $D + \Delta D$ the diffusion constant of another type of charged particle to be separated (with ΔD referring to the difference in diffusion constants);
- N_{cyc} the number of temporal cycles of the potential for a complete separation run;
- T_{tot} the total time of a complete separation run ($T_{tot} = P * N_{cyc}$ and $N_{cyc} = f * T_{tot}$);
- V_{drift} the drift velocity of charged particles in the electric potential; and
- 25 L_{tot} the total length of the separation lane.

First, the preferred embodiment for the method and device of this invention is presented.

Second, a method is presented for selecting operational and device parameters in an optimal manner. Third, exemplary alternative operation modes within the scope of this invention are

30 described.

5.3.1. An Embodiment Of The Invention

The section includes a discussion of the spatially and temporally varying potential of a method of this invention, of criteria for the operational method parameters, and of a

35 preferred model of the method in view of these criteria. Initially, it is assumed that $\Delta D \ll D$; subsequently, the case where $\Delta D \geq D$ is described.

THE POTENTIAL

Fig. 10 illustrates generally and schematically an electric potential, $V(x)$, as a function of distance along the separation axis, x , that is usable in this invention. This potential is spatially periodic with spatial period L . Alternatively, non-spatially periodic
5 potentials can be used in this invention. Every period of the potential should be eccentric with each minimum of the potential closer to the adjacent maximum in one direction along the separation lane. This direction is the direction of particle separation S . The separation between a minimum and its closest adjacent maximum is expressed by R , with $R < L/2$. For example, minimum 1003 is spaced a distance R , which is less than $L/2$, from adjacent
10 maximum 1002, but is spaced a distance $L-R$, which is greater than $L/2$, from adjacent maximum 1001. All the minimums are closer to the maximum adjacent in the direction S . This potential closely approximates the potential generated near the electrode pattern of device types I and II.

When $V_0 > 0$, the potential of Figs. 11A-D separates positively charged particles in
15 the direction S . In this case, negatively charged particles are transported through the device in the opposite direction, but are not necessarily separated. To separate negatively charged particles along direction S the polarity of the potential must be reversed, that is $V_0 < 0$. In this latter case, positively charged particles are transported through the device in the
20 opposite direction, but not necessarily separated. Advantageously, particles can be loaded in a loading zone at one end of the device and the device operated first with one polarity and second with the reverse polarity to sequentially separate particles of both charges. In the preferred embodiment specific to separations of DNA, however, it is envisioned that all the
25 particles have a negative charge.

The exact spatial configuration, $V(x)$, of the potential is not important to the operation of this invention. What is important is, first, that the potential consist of alternating potential maximums and potential minimums along the axis of separation. The wells are generally separated by distance L . Second, all the maximums and all the
30 minimums should be eccentrically placed in that each minimum should be closer to its adjacent maximum in the direction of separation than it is to its adjacent maximum in the direction opposite to the direction of separation. The distance between a minimum and the nearest maximum is generally R . Distances L and R are conveniently taken to characterize a region of the potential. The method of this invention is adaptable to any potential meeting
35 this constraint and separates particles in a direction of separation of a charge which is attracted into the wells. It is preferable that the potential be periodic having similar potential wells, and all subsequent discussion assumes spatial periodicity. Spatial periodicity is not required by the invention, however.

For greatest separation efficiency, that is for minimum separation time, it is preferable that the potential is homogeneous in directions transverse to the migration axis. The method and device of this invention functions, albeit at reduced efficiency, if the electric field vectors have components perpendicular to the axis of separation or, in other words, perpendicular to a disposition longitudinal to the direction of separation. The efficiency of the operation of the method is approximately $\cos(\Theta)$, where Θ is the angle of the electric field vectors relative to a disposition longitudinal to the direction of separation. Thus it can be appreciated that the method functions for nearly all relative directions between the electric field and the direction of separation. However, it is preferable that all electric field vectors be substantially longitudinal to the axis of separation. In this context, substantially longitudinal is preferably taken to mean that Θ is less than about 45-50° so that $\cos(\Theta)$ is therefore greater than about 0.5.

Moreover, in certain embodiments of the invention, it is possible to adjust operational parameters to minimize the effect of transverse inhomogeneities in the potential. For example, in an embodiment in which the electric potential is generated by electrodes adjacent to the separation lanes and in which the potential varies from a substantially off-state for a time t_{off} to a substantially on-state for a time t_{on} , for instance, in device type I or II, the potential may be inhomogeneous transverse to the migration axis due to electrode size and spacing in comparison to preferable separation lane widths. Since the electric potential decays away from electrodes, the potential wells are deepest closest to the electrodes. If the lane width is greater than the smallest inter-electrode spacing and if the electrodes do not entirely surround the channel, the potential well may be weak at the side of the lane furthest from the electrodes. However, this does not present a problem to this invention, because, first, t_{on} is optimally selected to attract and trap the charged particle at the electrodes and, second, t_{off} is optimally selected so that the particles diffuse at most a distance approximately equal to the inter-electrode spacing. Thus, since the particles to be separated remain in a region of relatively strong potential wells throughout the optimal operation of the device, potential inhomogeneity above the electrodes can be substantially neglected. Also, the electric potential may be perturbed near the separation lane walls. Again, this inhomogeneity can be substantially neglected because during operation because the charged particles remain where the potential wells are relatively strong.

The electric potential also varies temporally. All that is required is that the potential vary from a first strength, in which the particles are attracted to and trapped in the spatial potential wells, to a second strength, in which the particles are relatively free to diffuse in both directions, having a non-zero probability of diffusing into the nearest potential well. This probability can be quite small, 0.1% or less, or quite large, nearly 100%. It is preferably optimized to obtain the fastest possible separation. For convenience only and not

by way of limitation, subsequent description assumes that the temporal variation is periodic, with a period P and a frequency f , varying between an on-state and an off-state. For a time t_{on} , a potential $\pm V_0/2$ is applied, and for a time t_{off} , no potential is applied. Thus, during each cycle of operation of time T , the potential is on for time t_{on} and off for time t_{off} , with $t_{on} + t_{off} = T$ and $f = 1/T$.

Although, the method is modeled with a temporally periodic two state potential, potentials with other temporal variations can be used in the invention. First, the temporal variation need not be periodic. For example, the temporal period may change systematically as a separation progresses. Second, it is possible to include other states in the cycling. For example, a state can be included to focus the particles more tightly at the bottom of a well at the start of each cycle so as to obtain a smaller value for the parameter R . The potential may also vary continuously in time.

15 PREFERRED CONSTRAINTS ON METHOD PARAMETERS

When the potential is on, the potential wells should be sufficiently deep to attract and trap against thermal agitation the charged particles to be separated. This condition is met if V_0 is sufficiently large so that the following inequality is valid.

$$20 \quad \frac{V_0 Q}{k_b T} \gg 1 \quad (2)$$

(k_b is Boltzmann's constant.)

When the potential is off, particles of diffusion constant D in one well should have a finite probability, α_D , of diffusing a distance R in the direction of separation to the next potential well. The probability is advantageously chosen by optimizing t_{off} to obtain the fastest possible separation. In the preferred model for this invention, this condition is expressed as a relation between R and t_{off} given by

$$30 \quad \alpha_D = 1/2 \operatorname{erfc} \left(\frac{R}{\sqrt{4Dt_{off}}} \right) \quad (3)$$

"erfc" is the complementary error function.

35 Further, when the potential is off, particles of diffusion constant D should have a probability of diffusing backward a distance $L-R$ to the previous potential well that is preferably less than $\alpha_D/100$. In the preferred model for this invention, this results in the condition between R , L , and α_D .

$$\sqrt{4Dt_{off}} \leq R \ll L \quad (4)$$

5 The probability that particles diffuse a distance $L+R$ to the potential well beyond the next closest adjacent well is necessarily smaller than the probability they diffuse backward a distance $L-R$.

These conditions are easily met. For example, if $\alpha_D = 0.05$ and $R/L = 0.1$, then the probability of backward diffusion is vanishingly small, about 10^{-50} , and the probability of diffusing by more than one potential well is even smaller.

10

A PREFERRED MODEL

Various models can be constructed to aid in selecting operational parameters for the method of this invention and design parameters for the device of this invention. For example, the exact spatial and temporal structure of the electric potential generated by the electrodes actually used and the exact motion of charged particles in the separation medium subject to such a potential may be determined by the solving known differential equations of electromagnetism and particle motion. These equations can be numerically solved by standard methods (Press et al., 1992, Numerical Recipes in C, 2nd ed., Cambridge Univ. Press, New York (a cookbook of numerical procedures). Instead, it is preferred to construct an approximate model, which gives adequate results, and to optimize parameters based on experiments with actual devices. This preferred method results in adequate accuracy for operational and design parameters with less time and expense than an exact model.

25 The preferred approximate model describes the method and device of this invention as a random walk with drift. See, e.g., Wax, ed, 1954, Selected Papers on Noise and Stochastic Processes, Dover Publishers, New York. The random walk component is due to particle diffusion when the potential is off, and the drift component is imposed by particle trapping in potential minimums when the potential is on. The preferred model is herein described with reference to a preferred generally sawtooth-shaped electric potential, characterized by distances L and R , and with all particles initially trapped in one potential well in a loading zone. Then, the drift in the direction of separation during each cycle of the potential is $\alpha_D L$, where α_D is the probability for a particle of diffusion constant D to diffuse a distance R into the nearest potential well. Under the preferred parameter constraints, the probability that particles diffuse backward or forward by more than one potential well is negligible. The variance of particle position increases per potential cycle according to $(\alpha_D - \alpha_D^2)L^2$. The central limit theorem shows that the concentration profile of the particles as observed over many potential wells becomes a Gaussian distribution. See, e.g., Wax, ed, 1954, Selected Papers on Noise and Stochastic Processes, Dover Publishers, New York.

Therefore, after a time, t , that is after tf cycles, the Gaussian distribution of particle concentration has a peak, called $\langle x_D(t) \rangle$, given by:

$$\langle x_D(t) \rangle = tf\alpha_D L \quad (5)$$

5 The half-width of the Gaussian distribution of particle concentration, called $\delta x_D(t)$, is given by:

$$\left[\langle \delta x_D^2(t) \rangle \right]^{1/2} = \left[\langle x_D(t) L(1-\alpha_D) \rangle \right]^{1/2} \quad (6)$$

10 These expressions characterize the particle concentration across several potential wells.

Since the particles diffuse freely when the potential is off, α_D can be calculated as the fraction of particles that diffuse at least a distance R to the right during time t_{off} . According to standard diffusion theory, this is given by

$$15 \quad \alpha_D = \frac{1}{2} \operatorname{erfc}\left(R/\sqrt{4Dt_{off}}\right) \quad (7)$$

See, e.g., Wax, ed, 1954, Selected Papers on Noise and Stochastic Processes, Dover

20 Publishers, New York. In this expression, the complementary error function is defined by

$$\operatorname{erfc}(x) = (2/\sqrt{\pi}) \int_x^\infty dt \exp(-t^2). \quad (8)$$

Polynomial approximations for $\operatorname{erfc}(x)$ are found in Abramowitz et al., 1972, Handbook of
 25 Mathematical Functions, Dover Publishers, New York. Eqn. 7 for α_D assumes that the initial distribution of particles in each potential well is of very small width. In fact, the initial distribution of the particle density trapped in the bottom of each well when the potential has finite width, is on the order of the width of an electrode. However, this
 30 difference affects only the numerical predication of α_D . It does not affect the model, in particular, Eqns. 5 and 6, since all these require that α_D functionally depend in a known manner on the diffusivity, D .

This model demonstrates how particle species of different diffusivities are separated by this invention. First, eqns. 5 and 7 demonstrate that a species with a greater diffusivity is
 35 transported in the direction of separation more rapidly than a species with a lesser diffusivity. Eqn. 5 also demonstrates that the separation between species of different diffusivities grows linearly with time, t , or equivalently the number of cycles, N_{cyc} . Second, Eqn. 6 demonstrates that the widths of the concentration profiles of each species increase as $t^{1/2}$. Since the separation between concentration profiles for each species grows more rapidly

than the widths of the concentration profiles of any species, after a sufficient number of potential cycles the concentration peaks associated with species of different diffusivities becoming spatially separated in an observable manner.

Moreover, Eqn. 7 demonstrates how the time required for separation depends on
5 device feature size. The shape of the concentration profiles of the species and the rate of separation of the species are determined entirely by the probability parameter α_D . As this parameter, in turn, depends only on the argument of the complementary error function, a change in the feature size R can be balanced by a change in the time t_{off} that leaves this argument unchanged. Since R enters linearly and t_{off} enters as a square root, a 2X reduction
10 in the feature size permits a 4X reduction in the time required for a separation. The overall length of the device scales linearly with R (at constant R/L). Therefore, separations are increasingly rapid for sufficiently small device length. Thus, advances in microfabrication technologies can be applied to enhance directly the performance of the device by reducing
15 its feature size. The devices should remain larger than the previously discussed minimum, *i.e.*, an order of magnitude larger than the size of the particles to be separated.

Figs. 11A-D and 12A-E illustrate the operation of the invention according to this model. Fig. 11A-D illustrates the detailed concentration profiles of two species of particles, A and B, of differing diffusivities, species B having greater diffusivity than species A, in
20 two adjacent potential wells, generally indicated at 1101 and 1102. Position 1108, also labeled R , is the nearest potential maximum adjacent to minimum 1101. In Fig. 11A, electric potential 1107 is on and the particles are attracted to initial potential well 1101 and tightly trapped against thermal spreading according to Eqn. 2. Concentration profiles 1103 of species A and 1104 of species B are generally Gaussian-like in each well. In Fig. 11B,
25 the electric potential is turned off and the molecules diffuse at rates dependent on their diffusivities in both directions in the separation medium. Species A and B now have broader Gaussian-like concentration profiles, 1103 and 1104, with profile 1104 of species B being broader as it has greater diffusivity. Some of species A, profile 1103, and more of species B, profile 1104, diffuse beyond adjacent maximum 1108. In Fig. 11C, potential 1107 is turned
30 on again and the particles again are attracted and tightly trapped in wells 1101 and 1102. However, now those particles that diffused beyond maximum 1108 are trapped in well 1102 in concentration profiles 1105 of species A and 1106 of species B. These particles have drifted one well forward. More of species B than of species A is in well 1102. In Fig. 11D,
35 the potential is turned off again and both species diffuse outward from both wells. Due to the asymmetry of the potential, the concentration profile of the molecules has been selectively transported to the right with the species of greater diffusivity being transported faster.

Fig. 12A-E illustrates the operation of the model of the invention on a scale of many potential wells. These figures are generated from an exact calculation based on the random walk with drift model. The horizontal axis is along the direction of separation and includes 30 potential wells. The vertical axis represents the concentration of the two species of charged particles to be separated, species A and B. Species A, of lesser diffusivity, is represented by bars 1201, and species B, of greater diffusivity, is represented by bars 1202. The time t_{on} and t_{off} are chosen optimally according to methods to be described. Fig. 12A illustrates the initial condition in which both species are trapped in the first potential well only. Figs. 12B, 12C, 12D, and 12E show the concentration profiles of both species after 25, 50, 75, and 100 cycles, respectively. These profiles become increasingly Gaussian over many potential wells as the number of cycles increases, as required by the central limit theorem. From Figs. 12B-E, it is apparent that species B is transported to the right faster than species A and that both concentration profiles spread over time. It is also apparent the species are being separated since the concentration peaks are moving apart faster than the concentration profiles are spreading.

5.3.2. Choice Of Optimum Parameters

Optimal selection of method operating parameters and device design parameters depends on which characteristics of the particle separation are to be optimized. This section describes a method for minimizing the separation time according to the preferred model. It will be apparent that the same method can be applied to more realistic device models incorporating more structural details of the device, the potential, and particle transport. Alternately, in an analogous manner according to both the preferred model and more complete models, one of average skill in the art can optimize other separation characteristics, such as, for example, the spatial distance of separation.

The preferred operating parameters are chosen to minimize total separation time, which is determined by the potential cycle time, $t_{on} + t_{off}$. This section describes, first, the optimization of t_{on} and related parameters, and second, the optimization of t_{off} and related parameters.

Parameters selected by these methods are necessarily approximate. More accurate optimal parameters can be determined from the parameters herein determined by routine experimentation with actual devices. Further, an actual device need not be operated at the exactly optimal parameters determined according to any method. One of skill in the art will recognize that an actual device can be operated with parameters deviating slightly or substantially from the exactly optimal in order to accommodate, for example, the characteristics of the available equipment, inaccuracies in setting operating parameters, etc.

It is only preferable to operate a device near the determined parameters in order to achieve the optimums.

The methods described herein are capable of implementation as a computer program by routine translation into an appropriate computer language, such as C, Basic, Fortran, etc.

- 5 This computer program can command a general purpose computer system to perform the parameter selection methods described. Such a computer system can be, for example, an IBM or equivalent PC.

PREFERRED OPTIMIZATION OF t_{on} AND RELATED PARAMETERS

- 10 It is preferable to select operational and device parameters so that t_{on} is as small as possible. Here, first, relations relating t_{on} to relevant parameters are determined according to the preferred model, and second, these relations are used to determine optimal values for these parameters.

- 15 The time t_{on} is the time for a charged particle to drift in the direction of separation under the influence of the potential from a maximum in the potential to the subsequent minimum, a distance $L-R$. For example, in Fig. 10, t_{on} is the time for a particle to drift from 1001 to 1003. This time is given by:

$$20 \quad t_{on} = (L-R) / V_{drift}, \quad (9)$$

where V_{drift} is the drift velocity of a particle in the potential.

- Since the motion of a particle in the separation medium is over-damped, V_{drift} is proportional to the force from the electric potential times a friction coefficient, and is given
25 by

$$V_{drift} = \gamma Q \left(\frac{V_o}{L-R} \right). \quad (10)$$

- 30 Note that since the potential must be eccentric, $R < L/2$. The electric field E in the pertinent region of drift is $-V_o / (L-R)$. The friction coefficient, γ , is related to the diffusion constant by a fluctuation-dissipation theorem

$$\gamma = D / k_B T. \quad (11)$$

- 35 See, e.g., Wax, ed, 1954, Selected Papers on Noise and Stochastic Processes, Dover Publishers, New York. Combining these equations, t_{on} is given by

$$t_{on} = \frac{k_b T (L-R)^2}{QDV_o} \quad (12)$$

5 Therefore, according to Eqn. 12, given $L-R$, the minimum and hence most preferable t_{on} can be calculated. Further, V_o should be selected to be as large as possible consistent with the electric field remaining less than the breakdown field and electrolysis threshold of the separation medium used. The maximum electric field, E_{max} arises along the steeper side of the potential and is given by

$$E_{max} = \frac{V_o}{R} \quad (13)$$

15 Optimally, the device should be operated as close to the limiting field as possible. In this case, t_{on} is given by

$$t_{on} = \frac{k_b T (L-R)^2}{QDRE_{max}} \quad (14)$$

20

For example, in the case of water the limiting breakdown field is approximately 10^4 V/cm (Avallone et al. eds., 1987, *Marks' Standard Handbook for Mechanical Engineers*, McGraw-Hill, New York, pp. 15-19). Therefore, if $E_{max} = 10^4$ V/cm and $R = 1 \mu\text{m}$, then the maximum V_o is 1 V.

25

PREFERRED OPTIMIZATION OF t_{off} AND RELATED PARAMETERS

In order to optimally select t_{off} and related parameters it is necessary to specify what is meant by a successful separation of a particle type of diffusivity D from a type of
 30 diffusivity $D + \Delta D$. In this section, it is assumed that ΔD is much less than D . A preferred separation specification is that separation occurs when the difference between the exit times from the device of the concentration peaks of the two types of particles is at least as large as the spreading of the concentration peaks. In this case, the concentration profiles of the two types of particles can be experimentally distinguished. Alternatively expressed, separation
 35 occurs at that time, or number of cycles, when the difference in positions of the concentration peaks of the two types of particles is at least as large as the Gaussian spreading of the two peaks. For example, in Figs. 12A, 12B, and 12C the two concentration peaks would not be considered as separated according to this preferred specification.

However, in Figs. 12D and 12E the peaks would be considered as separated. Alternative more or less stringent separation specifications can be applied to select operational parameters. A less stringent condition might consider, for example, Fig. 12C as also separated.

5 According to this preferred separation specification, separation occurs at a time t_D given by:

$$\langle x_D(t_D) \rangle - \langle x_{D+\Delta D}(t_D) \rangle = \left[\langle \delta x_D^2(t_D) \rangle \right]^{1/2}, \quad (15)$$

10 Here, upon exiting the device after time t_D , the time required for a particle of diffusivity D to traverse the device, the concentration profiles of the two types of particles are separated.

From Eqns. 5 and 6, the separation condition can be written as

$$t_D f(\alpha_D - \alpha_{D+\Delta D}) L = \sqrt{t_D f \alpha_D L^2 (1 - \alpha_D)}, \quad (16)$$

where α_D is the probability that a particle of diffusivity D diffuses to the next potential well during t_{off} . α_D is given by Eqn. 7, which is repeated here for convenience.

$$20 \quad \alpha_D = \frac{1}{2} \operatorname{erfc}\left(R / \sqrt{4 D t_{off}}\right) \quad (17)$$

From Eqn. 16, the number of cycles, N_{cyc} ($= t_N f$), required to separate particles of diffusivity D from particles of diffusivity $D + \Delta D$ is given by:

$$25 \quad N_{cyc} = \alpha_D (1 - \alpha_D) / (\alpha_D - \alpha_{D+\Delta D})^2. \quad (18)$$

The difference $\alpha_D - \alpha_{D+\Delta D}$ can be approximated as $-\Delta D \partial \alpha_D / \partial D$ for sufficiently small ΔD . The total separation time is:

$$30 \quad T_{tot} = N_{cyc} (t_{on} + t_{off}) \quad (19)$$

The preferred optimum parameters are selected to minimize T_{tot} . Operational and device parameters have previously been selected to minimize t_{on} . T_{tot} depends on t_{off} both directly, through Eqn 19, and indirectly, since N_{cyc} depends on α_D which in turn depends on t_{off} . To select the optimum value of t_{off} , all these equations must be minimized together. This minimization is most easily done by standard numerical methods, for example, by systematically trying various values for t_{off} until a minimum is found. See, *e.g.*, Press et al.,

1992, Numerical Recipes in C, 2nd ed., Cambridge Univ. Press, New York. An example of the selection of an optimum t_{off} is given in Sec. 5.4 for the case of DNA fragment separation.

Once optimum t_{off} and N_{cyc} values have been selected, the total device length required for separation is given by

$$L_{tot} = \langle x_D(t_D) \rangle = N_{cyc} \alpha_D L. \quad (20)$$

The preferred optimum quantities selected depend on the spatial characteristics of the potential.

10

PREFERRED OPTIMIZATION OF L AND R

The preceding optimization of t_{on} and t_{off} assumed that R and L are fixed. If these lengths can be varied they should be selected in view of the previous optimum time parameter determinations. First, in view of Eqn. 14, L should be selected as small as possible. Second, in view of Eqn. 17, since a 2X reduction in R allows a 4X reduction in the t_{off} , R should be chosen as small as possible. Third, to have sufficient eccentricity of the potential wells, it is preferable that $R/L < 0.3$. And fourth, R and L are limited to be at least as large as the minimum dimensions permitted by a chosen fabrication technology. These conflicting requirements on R mean that for a chosen separation medium with a fixed E_{max} (V_o being varied) an optimum R exists.

In a preferable method for optimization, the separation time T_{tot} is minimized as a function of L and R , subject to the constraint that the resulting device sizes can be fabricated by the chosen fabrication technologies. An additional constraint is that for each value of L and R , it is optimum to select the applied potential, V_o , such that the electric fields are smaller than the breakdown field, E_{max} . For this value of V_o , the times t_{on} and t_{off} are selected in an optimum manner, as described previously, to arrive at an optimum T_{tot} for given L and R . Using multidimensional minimization techniques that are well-known to those of average skill in the computational arts (Press et al., 1992, Numerical Recipes in C, 2nd ed., Cambridge Univ. Press, New York) the optimum pair of L and R is then readily and preferably determined from this optimization problem.

In another method, the optimum R can be determined by the simultaneous minimization, within the technology allowed bounds, of Eqns. 14, 17, 18, and 19. This can be preformed by standard numerical techniques from Press et al. Alternatively, this minimization can be performed by the following simple search procedure. Pick an initial R at the minimum allowed bound and determine an optimum T_{tot} by the previous methods. Increase R by some fraction, say 5%, and repeat the determination of an optimum T_{tot} . For all determinations of T_{tot} , chose the maximum value of V_o that can be applied in the given

separation medium selected. Continue this iteration until a minimum value for T_{tot} is found, either at the lower bound on R or at an intermediate value of R . The preferred value of R is the one that minimizes T_{tot} . Having chosen R , L can be preferably determined so that R/L has a fixed value providing sufficient eccentricity of the potential wells. Preferably R/L is less than 0.3, and more preferably is approximately 0.1.

For all of the sample calculations below, the optimum values are assumed to be $R = 1 \mu\text{m}$ and $L = 10 \mu\text{m}$, unless specified otherwise. A potential with this periodicity is readily produced by common microfabrication techniques, such as those described in Sec. 5.5.

Finally, the separation medium should be chosen in order that the particles to be separated are suspended in the medium in a charged state and have differing diffusivities when suspended. The greater the difference in diffusivities, the more preferable is the medium. It is further preferable that the separation medium be chosen from among the otherwise suitable mediums to have a relatively high E_{max} and a relatively high electrolysis voltage compared to the other suitable mediums. Here, relatively high can be taken to be at least greater than the average values for the otherwise suitable mediums. These conditions permit a minimum t_{on} . Finally, it is preferable that the separation medium have low ionic strength to minimize screening of the potential wells.

5.3.3. The Case Of Widely Varying Diffusivities

The preceding sections have described a preferred model, and the determination of preferred operating and device parameters in view of the model, for the case where the particles to be separated have similar diffusivities. This invention is also applicable to mixtures which contain particles with widely varying diffusivities.

One mode of operation to separate such mixtures is to begin the with t_{on} and t_{off} at the short times optimal for separating particles of higher diffusivities. With these times, the more diffusive particles are rapidly separated. However, the particles of lower diffusivities, having much smaller α_D values, remain nearly stationary. After the more diffusive particles have been separated, the times t_{on} and t_{off} are increased to the larger values optimal for separation of the less diffusive particles. The less diffusive particles are then rapidly separated subsequently.

Another mode of operation to separate such mixtures is to use the longer t_{on} and t_{off} times appropriate for the less diffusive particles. With such longer times, the more diffusive particles have larger α_D values and may be able to diffuse more than one potential well in the forward direction as well as in the backward direction. The previous model assumed that particles to be separated either did not move or diffused at most one potential well in the forward direction during t_{off} . However, a similar model based on random walk with drift can

be constructed for the case in which some of the particles to be separated diffuse more than one potential well during t_{off} .

To construct such a model, define $\alpha_D^{(n)}$ as the probability that a particle with diffusivity D diffuses n potential wells during the time t_{off} for free diffusion. According to standard diffusion theory in a manner similar to that of Eqn. 7, these α 's are given by

$$\alpha_D^{(n)} = \frac{1}{\sqrt{4\pi Dt_{off}}} \int_{R^{-(n-1)L}}^{R+nL} dx \exp(-x^2/4Dt_{off}). \quad (21)$$

When the diffusivity, D , is relatively small compared to t_{off} , $\alpha_D^{(1)}$ rapidly approaches the value for α_D of Eqn. 7. In this case, $\alpha_D^{(1)}$ and $\alpha_D^{(0)}$ (the probability that the particles stay put) are the only non-zero $\alpha_D^{(n)}$, implying that the particle either stays put or diffuses forward by a single potential well spacing. However, when the diffusivity is relatively large, $\alpha_D^{(n)}$ for $n = -1, 2$, etc., can become important.

To model the invention in the case that particles can diffuse more than a potential well during t_{off} , redefine the parameter α_D as the effective probable diffusion distance

$$\alpha_D = \sum_{n=-\infty}^{\infty} \alpha_D^{(n)} n. \quad (22)$$

With this definition, the average position of the maximum of the Gaussian-like particle concentration profile, again called $\langle x_D(t) \rangle$, is given by

$$\langle x_D(t) \rangle = t f \alpha_D L \quad (23)$$

This is the same as Eqn. 5 of the previous model, where the particles were assumed to diffuse by one potential well at most.

The variance in the Gaussian-like particle concentration profile of particles of diffusivity D after total time t , again called $\delta x_D(t)$, is equal to the number of diffusion cycles in time t , that is tf , times the variance change $\langle \delta x_D^2 \rangle$ for a single cycle, which is in turn given by

$$\begin{aligned} \langle \delta x_D^2 \rangle &= \langle x_D^2 \rangle - \langle x_D \rangle^2 \\ &= \sum_{n=-\infty}^{\infty} \alpha_D^{(n)} n^2 L^2 - \alpha_D^2 L^2 \end{aligned} \quad (24)$$

Again, the preferred condition defining the occurrence of separation of particles of diffusivity D from particles of diffusivity $D + \Delta D$, where now ΔD is of the order of

magnitude of D or larger, is that distance between the concentration peaks must be at least as large as the half-width of the concentration profile. From Eqns. 23 and 24, this condition for separation is given by

$$N_{cyc} L(\alpha_D - \alpha_{D+\Delta D}) = \sqrt{N_{cyc} \langle \delta x_D^2 \rangle}. \quad (25)$$

These equations permit the same conclusion for this case, where ΔD is of the order of D or larger, as for the previous case where $\Delta D \ll D$. As previously, Eqns. 21 and 23 demonstrate that particles of greater diffusivity are transported more rapidly through the device than particles of lesser diffusivity and that the distance between the concentration peaks increases linearly with time. Eqn. 25 demonstrates that peaks of different diffusivities are separated. Again, separation occurs since the distance between the concentration profiles grows linearly with N_{cyc} while the width of the concentration peaks grows only as the square root of N_{cyc} . Further, because of the behavior of Eqn. 21, the separation time is decreased by 4X for every 2X reduction in the spatial scale of the potential.

Further, operational parameters optimizing t_{on} can be selected in a manner similar to the previous case. For example, N_{cyc} is given by

$$N_{cyc} = \frac{\sum_{n=-\infty}^{\infty} \alpha_D^{(n)} n^2 - \alpha_D^2}{(\alpha_D - \alpha_{D+\Delta D})^2} \quad (26)$$

This corresponds to the form of Eqn. 18. Therefore, the t_{off} that minimizes T_{tot} can be obtained by numerical minimization of Eqns. 19, 21, 22, and 26, similarly to the previous case in which the particles diffused at most by one potential well during t_{off} .

Thus mixtures of particles with widely varying diffusivities can be separated by the method even if operational parameters are chosen so that the more diffusive particles diffuse more than one potential well during t_{off} . In one mode of operation for separating such mixtures, the cycling times t_{on} and t_{off} can be first optimized for rapid separation of the more diffusive particles, and then gradually increased to larger values optimized for the separation of less diffusive particles. In a second mode of operation, the cycling times can be optimized to separate the largest fragments, still providing an adequate separation for the smaller fragments.

5.3.4. Alternate mode of operation: multiple states

Another mode of operation consists of cycling the potential through three states instead of two states. These three states include the following steps:

1. turning the potential on;
- 5 2. briefly reversing the potential one or more times during the on-condition; and
3. allowing free diffusion.

Briefly reversing the potential one or more times during the on-condition can be effective in reducing the electrostatic screening from an ionic double layer formed by small, highly mobile counterions attracted to the potential wells or electrodes. These counterions
 10 can be displaced and the double layer minimized by interspersing with t_{on} several rapid pulses in which the polarity of the potential is reversed. Preferably, the period of reversal is sufficiently small such that, although there is substantially no effect on the distribution of the larger, less mobile particles, the highly mobile, smaller counterions are displaced out of the
 15 potential wells or in a reverse direction from the electrodes. This is accomplished by satisfying the inequality $t_{pulse} \ll t_{on}, t_{off}$.

Another three-state mode of operation uses a third state with a sharp, substantially symmetric V-shaped potential centered at the bottom of each potential well, such as may be created by a third electrode between the two sets of electrodes present in device types I and
 20 II. The electrodes of this embodiment are located at relative positions $-R$, 0 , and R , with a periodicity of L . In the first state, which lasts for time t_{on} , the electrodes at relative position 0 are charged to $V_o/2$ and the electrodes at relative position R are charged to $-V_o/2$. In the third state, which lasts for time t_{off} , the electrodes are all uncharged and the particles diffuse freely. These first and third states are identical to the two states of the mode of operation
 25 described in previous sections. In the additional, third, middle state, the electrodes at relative positions $-R$ and $+R$ are charged to $+V_o/2$, and the middle electrodes at relative positions 0 are charged to $-V_o/2$. This creates a narrow V-shaped potential well, which tightly localizes the particles.

30 This is useful because it can provide a stronger and narrower trap for particles at the bottom of a potential well with steep walls on either side. This will produce a density distribution in each well in each potential cycle closer to the preferable vanishingly thin distribution.

5.3.5. Alternative mode of operation: temperature ramp

35 A further alternative mode of operation involves ramping or changing the temperature of the device during the course of operation, preferably increasing the temperature as transport and separation occurs. This mode is advantageous for separating groups of particles of widely varying diffusion coefficients and can be combined with the modes described in Section 5.3.3. At lower temperatures, since particles, such as molecules,

typically have a lower diffusivity (see, e.g., Eq. (27)) and a containing fluid typically has a higher viscosity, such lower temperatures are adapted to transport and separation of more mobile particles and molecules. On the other hand, at higher temperatures, since particles typically have a higher diffusivity (see, e.g., Eq. (27)) and a containing fluid typically has a lower viscosity, such higher temperatures are adapted to transport and separation of less mobile particles and molecules. According to this alternative mode of operation, device temperature would be lower for an initial time interval in order that more mobile particles in a sample can be separated and then would be higher for a later time interval in order that less mobile particles can be separated.

In more detail, the temperature can be ramped or increased in two or more steps or can be increased continuously. The size of the temperature increase can be chosen in order that the mobility of the less mobile species changes by a factor of approximately 1.2, 1.5, 1.8, 2.0, or more. For example, in the case of single-stranded DNA separation, the temperature can be increased from just above the freezing point of the solvent to just below its boiling point. Specifically, for aqueous or primarily aqueous solutions, the temperature could be increased from approximately 4°C to approximately 97°C. In this manner, for example, DNA molecules of 50 and 51 base pairs can be separated from, for example, DNA molecules of 300 and 301 base pairs in the same sample. The temperature ramp or change can be performed by the previously described thermal control module.

Further alternative modes of operation can be realized by dynamically modulating other conditions that determine the rates of particle transport. These conditions include gradient elution or solvent programming [D.A. Skoog, F.J. Holler, and T.A. Nieman, 1998, *Principles of Instrumental Analysis, 5th ed.*, Saunders College Publishing]. In general, any method or technique applicable to improve chromatographic separation can also be applicable to the separation device of this invention.

5.3.6. Alternative mode of operation: voltage ramp

An additional alternative mode of operation involves the ramping or variation of the potential of the device during the course of operation. In this embodiment of the invention, a cycle time is used for the potential that is too fast for the largest molecular fragments to follow, but is slow enough that the smaller fragments can follow. In such a case, separation is effected as a result of the different response to the voltage ramping by the differently sized fragments.

More specifically, the separation is effected by initially varying the potential with a high drive frequency. The frequency is set initially to optimize the separation of the smallest, fastest-moving particles. This frequency is gradually reduced, either continuously or discretely. This causes the smallest fragments to move the length of the chip, followed by

increasingly larger fragments. Since similarly sized fragments will respond to the voltage variation similarly, separation is effected. The rate of frequency reduction could be optimized for the separation desired given the diffusion rates of the species to be separated. There is no reliance on the isotropic diffusion of the molecules to be separated, although the separation is still based on molecular velocity in the carrier fluid, which is size and mass dependent.

In this approach, adjacent pairs of electrodes are not connected together. Rather, the voltage is swept down the length of the chip in a traveling wave. The larger, more massive fragments are unable to follow the wave whereas the smaller, less massive fragments are. Once the smaller fragments exit the detection region of the chip, the frequency of the traveling wave is lowered so the slightly larger fragments can follow it. This is repeated until all the fragments of interest are detected.

For example, consider a mixture of DNA fragments in the range from 20 to 200 bp and a device with dimensions r equal to $0.1\ \mu\text{m}$, L equal to $1.0\ \mu\text{m}$, and a voltage gap of $0.1\ \text{V}$ in the on-state. In a specific embodiment, one begins at a frequency between $1\ \text{kHz}$ and $10\ \text{kHz}$, most preferably with an on-time of $1.5 \times 10^{-4}\ \text{sec}$ and an off-time of $2.2 \times 10^{-5}\ \text{sec}$, and continues for $1.4\ \text{sec}$ in order to separate a 20mer from a 21mer ($D = 8.14 \times 10^{-7}\ \text{cm}^2/\text{sec}$ and $7.91 \times 10^{-7}\ \text{cm}^2/\text{sec}$, respectively, used for calculations). Most preferably, one switches to an on-time of $5.5 \times 10^{-5}\ \text{sec}$ and an off-time of $6.5 \times 10^{-5}\ \text{sec}$ and continues for $100\ \text{sec}$ in order to separate a 200mer from a 201mer.

5.4. Application to DNA Separation

An important application of the invention is to separate biopolymers (including biopolymer fragments), in particular, nucleic acids such as DNA (*e.g.*, cDNA, genomic DNA, synthetic DNA) and RNA. This application is possible because the diffusivity of DNA depends almost entirely on the number of nucleotides in the DNA molecule. There is an additional insubstantial dependence of the diffusivity on the total base composition, *i.e.*, for dsDNA the A+T to G+C ratio.

The required separation resolution depends on the application of DNA separation, ranging from a resolution of single base pairs to a resolution of as great as 10% of total DNA length or more. For example, for DNA sequencing, perhaps the more familiar application, DNA generally must be separated with a single base or base-pair resolution. Thus, aliquots of DNA generated by standard sequencing reactions (*e.g.*, F. Sanger et al., 1977, *Proc. Natl. Acad. Sci. USA* 74:5463; M. Maxam et al., 1977, *Proc. Natl. Acad. Sci. USA* 74:560) can be subjected to the separation methods of the invention. Another application of DNA separation, called sizing, requires a resolution of only $\pm 5\%$ or $\pm 10\%$ of the fragment length.

Sizing is used to produce quickly a pattern, or fingerprint, of the sizes in a DNA mixture, such as might be generated for a RFLP determination, genotyping, linkage analysis, microsatellite analysis and other fragment analysis application.

5

5.4.1. Diffusivity of DNA

The diffusion constants of ssDNA and dsDNA, used for selection of operating and device parameters, can be estimated from Stokes' law or obtained from experiment. The Stokes' law diffusion constant for particles is given by

$$D = k_B T / 6\pi\eta\Gamma, \quad (27)$$

10

in which T is the temperature, η is the viscosity of the separation medium (for example, for water, 0.01 gm/cm sec), and Γ is an effective particle radius (Doi et al., 1986, *The Theory of Polymer Dynamics*, Clarendon Press, Oxford, p. 300).

15

For a spherical particle, such as denatured ssDNA, Γ is identified as the radius of gyration. Scaling arguments relate the contour length of a polymer like ssDNA to its radius of gyration. In general, $\Gamma \sim N^\gamma$, where the exponent $\gamma \approx 0.6$ (see, e.g., Doi et al., *supra*). For a long cylinder with length $a \gg$ diameter b , such as dsDNA, one finds that $\Gamma \approx a / \ln(a/b)$. In dsDNA with N base pairs, the Stokes' Law diffusion constant with this approximation is given by:

20

$$\frac{\ln(a/b) k_B T}{3\pi\eta a} = (1/N) \ln(0.3N) \times 1.5 \times 10^{-5} \text{ cm}^2/\text{s} \quad (28)$$

25

in which diameter $b = 10 \text{ \AA}$ and length $a = 3N \text{ \AA}$. The temperature T is assumed to be 298°K throughout.

An experimental expression for the diffusion constant is preferable and is used throughout the following. The diffusivity of dsDNA at room temperature in water is given experimentally approximately by

30

$$D_{dsDNA} = 1.14 \times 10^{-6} N^{-1} \text{ cm}^2/\text{s} \quad (29)$$

in which N is the number of base pairs (Weast, ed., 1987, *Handbook of Chemistry and Physics*, Chemical Rubber Publishing Co, Boca Raton, FL, p 117). Observationally, the inverse dependence on N dominates the relatively weak $\ln(N)$ term in Eqn. 31. For ssDNA, the diffusivity is theoretically assumed to be given by

35

$$D_{ssDNA} = 1.14 \times 10^{-6} N^{-0.59} \text{ cm}^2/\text{s} \quad (30)$$

The scaling with N is derived from Stokes' Law, which predicts that D depends on the inverse of the effective radius. The effective radius is derived from considering that ssDNA diffusion resembles a self-avoiding walk, in which the effective radius depends on the number of bases as $N^{0.59}$. See, *e.g.*, Doi et al., *supra*.

5

5.4.2. Optimal Selection Of t_{on} And t_{off}

PREFERABLE DETERMINATION OF AN OPTIMUM t_{on}

t_{on} , the time required to attract DNA fragments in the potential wells, can be
 10 determined by combining Eqns. 12 or 14 for t_{on} with Eqns. 29 and 30 for the diffusion constant of DNA. The charge on DNA, Q , is $-N|e|$ for ssDNA and $-2N|e|$ for dsDNA, where N is the number of bases or base pairs and $|e|$ is the magnitude of the electronic charge. With V_0 in volts and $(L-R)$ in μm , t_{on} is given by:

$$15 \quad t_{on} = \frac{(L-R)^2}{(V_0/2) N^{0.41}} \times 1.1 \times 10^{-4} \text{ sec, ssDNA;} \quad (31)$$

$$20 \quad t_{on} = \frac{(L-R)^2}{(V_0/2)} \times 5.6 \times 10^{-5} \text{ sec, dsDNA;} \quad (32)$$

If V_0 is selected as the breakdown voltage for water, then t_{on} is given by:

$$25 \quad t_{on} = \frac{(L-R)^2}{RN^{0.41}} \times 2.3 \times 10^{-4} \text{ sec, ssDNA; and} \quad (33)$$

$$t_{on} = \frac{(L-R)^2}{R} \times 10^{-4} \text{ sec, dsDNA,}$$

30

with L and R in μm .

Table 2 shows t_{on} in seconds for a device with $L = 10 \mu\text{m}$, $R = 1 \mu\text{m}$, and $V_0 = 1 \text{ V}$. For the preferred most rapid separation, t_{on} should be chosen to be as small as possible.

35

TABLE 2

Fragment Size	t_{on} for ssDNA (secs)	t_{on} for dsDNA (secs)
10	0.0071	0.0091
100	0.0028	0.0091
500	0.0014	0.0091

5

The time t_{on} scales linearly with the spatial scale of the potential Φ and L). It scales differently with N for ssDNA and dsDNA. For ssDNA, since the driving force increases linearly with the molecule length, but the diffusivity decreases less rapidly, t_{on} is a decreasing function of the molecule length. For dsDNA, since the dependence of the driving force on the molecule length exactly cancels the dependence of the diffusivity on the fragment length, t_{on} is independent of molecule length.

15 PREFERABLE DETERMINATION OF AN OPTIMUM t_{off}

Preferably, an optimum t_{off} is selected to minimize the total separation time T_{tot} . T_{tot} is given by combining Eqn. 19 for T_{tot} with Eqn. 18 for N_{cyc} :

$$T_{tot} = (t_{on} + t_{off}) \alpha_D (1 - \alpha_D) / (\alpha_D - \alpha_{D+\Delta D})^2. \quad (34)$$

20

The dependence of α_D on D and t_{off} is given by Eqn. 7, which is repeated here:

$$\alpha_D = \frac{1}{2} \operatorname{erfc}\left(R / \sqrt{4 D t_{off}}\right) \quad (35)$$

25

D depends on N , the number of bases or base pairs in the DNA to be separated, according to Eqns. 32 and 33.

To minimize T_{tot} , the parameter t_{off} is systematically varied to obtain a minimum value for T_{tot} . The parameter t_{off} is optimally selected to be that value minimizing T_{tot} .

30

An exemplary program in the C language for calculating the parameters of DNA separation for this invention according to these relations, in particular Eqns. 33, 34, and 35, is set forth in Sec. 8. The input comprises the lengths R and L , the lengths N and $N + \Delta N$ for the DNA molecules to be separated, and a choice between ssDNA and dsDNA. The voltage V_0 is selected automatically to be the maximum consistent with the breakdown field of water and the over potential at which electrolysis of water occurs. The program can be changed for values of these parameters appropriate for other separation mediums. The program systematically varies t_{off} to find the optimum T_{tot} . The output comprises optimum operating conditions t_{on} and t_{off} and further details of the operation, including N_{cyc} and T_{tot} . The output also comprises a file containing the values of these quantities in a range about the optimum.

35

This program can be compiled and executed on any computer system containing a C language compiler and run-time system. One skilled in the art can translate this program into other similar languages for execution on computer systems having such languages.

Fig. 13 illustrates an example of the optimum selection of t_{off} by the use of the program. T_{tot} is numerically evaluated in terms of t_{off} for a potential with $L = 10 \mu\text{m}$ and $R = 1 \mu\text{m}$ in which a dsDNA molecule of length 100 is to be separated from one of length 105 in an aqueous separation medium. Fig. 13 shows the resulting graph of the relation between these two quantities. From Fig. 13, it is evident that the optimum choice for t_{off} is 0.10 sec, which yields a optimum total separation time of 6.8 min.

Table 3 presents the results of similar optimizations for a variety of DNA molecule lengths and required separation resolutions. In all cases the separation is in an aqueous medium with a potential having $L = 10 \mu\text{m}$ and $R = 1 \mu\text{m}$.

TABLE 3

DNA fragment	T_{tot}/min	L_{tot}/cm	t_{off}/sec	N_{cvc}	α_N
ssDNA					
$N = 10, \Delta N = 1$	0.48	0.083	0.0053	2.3×10^3	0.036
$N = 100, \Delta N = 1$	71.	4.4	0.015	2.3×10^5	0.019
$N = 100, \Delta N = 10$	0.85	0.057	0.016	2.7×10^3	0.021
$N = 500, \Delta N = 25$	7.0	0.18	0.038	1.1×10^4	0.017
$N = 500, \Delta N = 50$	1.9	0.052	0.039	2.8×10^3	0.019
dsDNA					
$N = 10, \Delta N = 1$	0.32	0.028	0.013	8.7×10^2	0.032
$N = 100, \Delta N = 1$	150	1.48	0.098	8.5×10^4	0.017
$N = 100, \Delta N = 10$	1.9	0.021	0.11	9.9×10^2	0.022
$N = 500, \Delta N = 25$	32.	0.067	0.50	3.8×10^3	0.018
$N = 500, \Delta N = 50$	9.0	0.020	0.52	1.0×10^3	0.020

Fig. 14 graphically presents a summary of a large number of such optimization calculations. The horizontal axis of the graph indicates the desired separation resolution, expressed as a percentage of the molecule length. The vertical axis indicates the required total separation time, T_{tot} , in minutes. The graphs indicate the separation times required for two molecule lengths, 100 and 500, for ssDNA and dsDNA. Again, all separation are in an aqueous medium with a potential having $L = 10 \mu\text{m}$ and $R = 1 \mu\text{m}$.

It is evident from Fig. 14 that separations are much more rapid for sizing, requiring only a 5-10% resolution, than for sequencing, requiring a 1% or less resolution. From Fig. 14, a factor of 10 change in the required resolution leads to a factor of 100 change in T_{tot} . For example, separation of molecules of length 100 with a resolution of 5 bases (5%) can be

performed 25 times more rapidly than the separation of molecules of length 100 with a resolution of 1 base (1%). Therefore, it can be clearly appreciated that rapid sequencing and extremely rapid sizing of DNA are possible with this device. Reducing the device size and increasing the diffusivity, for example, by changing the separation medium or by increasing temperature, shortens separation times for the device.

5.4.3. Eccentricity Of The Potential

This section describes an exemplary demonstration that a more eccentric potential is preferable for faster separation times. Optimum separation parameters are calculated for potentials with fixed a periodic length, L , of 10 μm , but with a varying R , the distance between the potential well and the nearest adjacent maximum. A smaller ratio R/L means the potential minimum is more eccentrically placed in each potential well. Table 4 presents the results of the calculations performed for separation of ssDNA fragments of length 100 with single base resolution (1%) in an aqueous medium. In all cases, we use $V_o = 2$ V.

TABLE 4

$R/\mu\text{m}$	$T_{\text{tot}}/\text{min}$	L_{tot}/cm	$t_{\text{off}}/\text{sec}$	N_{cyc}	α_N
2	246.	4.0	0.058	2.5×10^5	0.016
1	66.	4.2	0.015	2.4×10^5	0.017
0.5	21.	4.9	0.0041	2.3×10^5	0.022
0.25	9.5	6.5	0.0012	2.0×10^5	0.032

The performance of the device increases, that is the total separation time decreases, as the ratio R/L decreases.

5.5. Microfabrication Of Device Types I And II

A device operating according to a method of this invention may be of any physical size appropriate to the separation application and consistent with the previously described minimum sizes. In the preferred embodiment, where the device separates charged biopolymer fragments as rapidly as possible, the physical size is generally preferred to be as small as fabrication technologies permit and the intended separation medium allows. In this section, exemplary fabrication methods using standard microfabrication technologies are presented for device type I and II that are suitable for an aqueous separation medium with an applied potential difference of approximately 2 volts. These methods are exemplary, as this invention includes devices of other dimensions and fabrication according to other technologies.

The size of the exemplary device of types I and II is approximately 1 cm to 10 cm along the separation axis and approximately 1 cm to 10 cm transverse to the separation axis. The channels in the device are approximately 30-50 μm wide, 10 μm deep, and spaced apart every 100 μm , with a separation of approximately 50 μm between adjacent channels. The electrodes of each plurality are spaced apart approximately 20 μm , *i.e.*, $L = 20 \mu\text{m}$, and are approximately 0.8-1 μm wide. The electrodes of each plurality are relatively displaced by approximately their width, *i.e.*, $R = 0.8\text{-}1.0 \mu\text{m}$. The electrodes of each plurality are connected to electrode pads at the edges of the device for linking to an external voltage source.

Except where noted, the following microfabrication plans apply equally to devices of both types. The methods described are standard in the microfabrication art (Sze, 1988, VLSI Technology, McGraw Hill, New York).

5.5.1. Substrates

A preferred substrate for the device is silicon. Alternative substrates include glass, fused silica, borosilicate, quartz, pyrex, and plastics such as polymethylmethacrylate, polycarbonate, polystyrene, polyimides, etc. The dimensions of the glass substrate are approximately 1-10 cm x 1-10 cm, with a thickness of 1-5 mm. A suitable source for a glass substrate is a microscope slide of soda lime glass, for example a 75 x 50 x 1 mm slide (Fisher Scientific catalog No. 12-550C).

Prior to all other microfabrication steps, the substrate should be cleaned. For glass substrates, this can be done by immersing the substrates in a hot bath of $\text{H}_2\text{SO}_4/\text{H}_2\text{O}_2$, rinsing in H_2O for 10 min, rinsing again with H_2O , and drying in an oven at $\approx 150^\circ\text{C}$ for 10 min to remove adsorbed water.

5.5.2. Electrode Fabrication

The electrodes for the device can be fabricated from various metals. Preferred metals are Al, Ag, Au, and Pt. Al is advantageous in that readily available CMOS foundries can be used, and disadvantageous in that it is more susceptible to electrolytic decomposition than is a noble metal. Other metals, including Ti, Ni, Zn, Ru, Pd, Ta, and W, as well as doped polysilicon can be used for device electrodes. Alternative electrode fabrication methods for these metals are described: a first method using etching and suitable for all the preferred metals; a first alternative suitable for Pt, and a second alternative suitable for Au.

Prior to electrode fabrication with the first method, or with the first alternative method, a patterning mask is produced for photolithography. Fig. 15 illustrates an exemplary mask. Electrodes, as at 1501, 1502, 1503 and 1504, are disposed substantially transversely to separation axis S. Each electrode is approximately 1 μm wide. The

electrodes form two pluralities, electrodes of each plurality being connected to one of electrode pads 1505 and 1506. These pads are macroscale, approximately 0.1 mm, and serve as contact points to an external voltage source. Electrodes of each plurality are periodically spaced with distance L and are displaced with respect to each other with distance R . A mask of these dimension is readily constructed with standard
5 microlithographic technology. For example, a suitable mask is obtained by selectively removing chrome deposited onto a quartz surface. The chrome is removed, for example, using computer aided design that provides input for a pattern generator. If obtainable, smaller feature sizes are preferable; the sizes used here are exemplary.

10 A first method for fabricating electrodes begins with depositing a uniform 1 μm thick layer of the selected metal on the side of the substrate which is to carry the electrodes. The metal layer can be from approximately 100 nm to approximately 1 μm , and is preferably approximately 300 nm. The metal can be deposited by, for example, physical vapor
15 deposition, chemical vapor deposition, or sputtering. Then a positive photoresist is spin-coated on top of the layer of metal, and is stabilized by soft-baking. The features on the mask are transferred to the photoresist by irradiation with UV light, and the unprotected regions exposed to the light are dissolved by an appropriate solvent. The surviving photoresist is fixed in place by hard-baking at a high temperature.

20 The electrodes are generated by etching the region of metal unprotected by photoresist. For Al electrodes, the etching can be accomplished by exposing the surface to Cl_2 vapor. The Cl_2 molecules react with Al atoms on the surface to produce AlCl_3 , which is volatile and leaves the surface. Wet etching is not preferable for Au and Ag electrodes
25 because undercutting will destroy the μm -scale electrodes. These features are preferably etched using Ar^+ ion milling. In this method, Ar^+ ions from an Ar radio-frequency plasma are accelerated into the surface and cause etching by physical bombardment. Milling enables the electrodes to be produced with straight side-walls, preserving the mask dimensions.

30 After etching, the remaining photoresist is removed from the surface of the substrate and the surface carrying the electrodes is cleaned for subsequent processing.

A first alternative method suitable for Pt electrodes uses standard micromachining technologies. This method is also suitable for Ru and Pd electrodes. The fabrication begins with the deposition of a 10 nm thick Ti layer using an evaporation system. This Ti layer acts
35 as an adhesion layer between the subsequent Pt layer and the glass. Next, a 100 nm thick layer of Pt is deposited on top of the Ti using an Ar ion sputtering system. The electrodes are defined in the metal layers using photolithography and etching. This process begins with spin-coating a photoresist on top of the Pt and exposing the photoresist with UV light

through the photolithography mask that has the electrode pattern on it. The exposed areas of the photoresist can then be dissolved away in a developer to leave the unexposed regions that define the electrode pattern. The photoresist will protect the areas of the metal to be retained; the rest of the metal is removed using ion milling. In the milling process,
5 positively charged Ar^+ ions are electrically accelerated to impinge on the surface of the metal and physically erode the layers. Once this etching is completed, the photoresist is dissolved with acetone to leave the finished electrodes.

A second alternative method suitable for Au uses micro-contact printing (μCP) (Xia et al., 1995, *J. Am. Chem. Soc.* 117:3274-3275; Jackman et al., 1995, *Science* 269:664-666).
10 Instead of a photolithography mask, an elastomeric stamp made according to an identical pattern of the same dimensions is used. Figs. 16A-B illustrate exemplary patterns. The stamp can be fabricated from polydimethylsiloxane. As before, Au is deposited to a thickness of 1 μm on the surface of the substrate using standard methods. Next, the
15 elastomeric stamp is wetted with an alkanethiol and pressed against the gold surface. A suitable alkanethiol is $\text{CH}_3(\text{CH}_2)_{15}\text{SH}$ (Kumar et al., 1994, *Langmuir* 10:1498-1511). Controlled spreading of the patterned self-assembled alkanethiol monolayer on the gold surface can be achieved by performing the printing under water, which has the further benefit of shrinking the feature size in a predictable manner. The stamp and substrate are
20 removed from the water and dried using N_2 gas, and then the stamp is removed from the substrate. Unprotected gold is removed by immersion in a cyanide solution (0.1 M KCN, 1 M KOH) with vigorous stirring using air or oxygen as an oxidant (Kumar et al., *supra.*). After a good rinse, the alkanethiol is removed from the surface to yield the pluralities of
25 gold electrodes.

5.5.3. Device Type I Channel Fabrication

Device type I channels can be fabricated by wet-etching of a glass substrate. Prior to channel fabrication, a photolithography mask must be constructed. Fig. 16A illustrates an
30 exemplary mask. The channels are defined by transparent bands, as at 1601 and 1602, on the otherwise opaque mask. Width C of each band is the desired channel width minus any expected undercutting during the etching process. For the process described, since the expected undercutting is 8-10 μm , 40 μm wide bands produce channels of the desired 55-60 μm final width. Width W between the bands is the desired channel spacing plus any
35 expected undercutting.

Fig. 16B illustrates an alternate channel geometry. Here channels, such as channel 1606, converge from a wide spacing at a loading zone, indicated generally at 1603, to a narrow spacing at a detection zone, indicated generally at 1605. The wide spacing in loading zone 1603 allows the channels to accommodate injection ports, such as port 1604, of

a diameter greater than the desired spacing between the channels. Although Fig. 16B illustrates piece-wise linear channels, alternative channel geometries, for example curvilinear, are adaptable to this invention.

A suitable photolithography mask may be fabricated by selectively removing chrome deposited onto a quartz surface. The chrome is removed, for instance, using a computer-generated design that serves as input to a pattern generator.

Channel fabrication begins with spin-coating a positive photoresist onto the glass substrate. A suitable photoresist is generated by exposing the substrate to hexamethyldisilazane vapor for 5 min, spin-coating with photoresist (Microposit S1400-31, Shipley, Newton, MA), and stabilizing the photoresist by heating at 90°C for 0.5 hr. The mask is aligned over the coated glass substrate and the pattern is imprinted on the photoresist using UV light. The regions of photoresist exposed to the UV light are dissolved away (1:1 mixture of H₂O and Microposit developer concentrate, Shipley), and the surviving photoresist is fixed by baking at 150°C for 1 hr.

Alternately, the channels may be defined in the glass substrate using a Cr layer. This process starts with evaporating a 100 nm thick Cr layer onto the glass. The Cr layer over the lanes to be fabricated is then removed using photolithography and etching.

Next, the unprotected areas of the substrate are wet-etched by exposing the surface of the glass chip to an aqueous NH₄/HF etching solution (1:1 mixture of BOE 5:1 and BOE 10:1, J.T. Baker, Phillipsburg, NJ). Etching for 20 min produces channels 10-15 nm deep, and undercuts the photoresist 8-10 µm on each side. The 40 µm feature size on the patterning mask therefore generates channels of the exemplary width of 55-60 µm. After etching, the photoresist or Cr layer is removed from the substrate, for example, in the case of glass, by cleaning with hot H₂SO₄/H₂O₂ as previously described.

5.5.4. Device Type II Channel Fabrication

Channels for device type II are preferably fabricated on top of the electrodes.

Alternatively, they can be fabricated on top of the other substrate. A suitable photolithography mask is first fabricated. Such a mask is generally similar to that for the device type I channels with the three exceptions. First, the mask defines two channel walls banding each channel. Second, the channel wells are defined by transparent bands with the remainder of the mask being opaque (a negative mask). Third, as no undercutting is expected in this method, the mask dimensions should exactly match the intended channel and channel wall dimensions.

Then, the channels are fabricated by first spin-coating a UV-sensitive polyamide solution on top of the surface of the substrate to a depth of approximately 10 µm. The polyamide remains in place in the regions which are exposed to UV light, requiring that the

photolithography mask be a negative image. The mask pattern is imprinted on the polyamide photoresist by illumination with UV light. The region of the polyamide layer under clear portions of the mask is stabilized by cross-linking due to the UV radiation. The remainder of the polyamide layer is dissolved with a suitable developer. The cross-linking
5 forms a straight side-wall which is preserved during the developing and curing stages. Next, the glass chip is hard-baked at approximately 150°C to set the polyamide layer. This completes the microfabrication of the channels.

5.5.5. Injection Port Fabrication

10 Injection ports may be fabricated in the substrate that does not carry the electrode pattern if desired. Holes for the injection ports can be fabricated by drilling either by a laser or a diamond tipped drill bit or ultrasonic drilling. Preferably, the drilled holes are sized to permit the injection of sample with a micropipette tip, so 500 µm is an adequate size. Since
15 the preferable size of the injection ports is 5 to 10 times the preferable spacing between the channels, the converging channel pattern of Fig. 16B is preferable to the straight pattern of Fig. 16A for closely spaced channels in the migration and detection regions.

5.5.6. Fusing The Substrates

20 In order to create closed separation lanes in the device, the substrate with the channels and the other substrate must be bonded together. First, both sides are cleaned thoroughly and then are brought into contact. For device type I, the temperature is steadily increased to the annealing temperature of approximately 500-600° C, where it is held for a few hours to ensure good bonding of the surfaces. For device type II, a flat silica plate is
25 fused to the polyamide surface of the channels at a lower temperature of approximately 200°C.

5.6. Description of Electrode Configurations

30 A main advantage of the electrode-design aspect of the invention is that it provides a means to collect a substantial concentration of charged particles to a confined region prior to separation; this collection meanwhile reduces the concentration of the particles throughout the rest of the device. This advantage is still available when the device is operated with the following steps: first, supplying the charged particles uniformly throughout the device;
35 second, sealing the device; third, focusing the charged particles to said confined region; and, last, activating separation.

A separation device can be constructed from a number of adjacent cells for holding two or more species of charged particles, together with a mechanism for transporting from each cell to its (right-hand) adjacent cell a species-specific proportion ($\alpha[\text{species}]$) of the

amount of each species within the cell. Operated repeatedly, starting from an initial condition in which substantially all of each species is in the left hand cell, transport and separation of the species will occur, directed to the right. The so-called Feynman ratchet (R.P. Feynman et al., 1966, *The Feynman Lectures on Physics, Vol. 1*, Addison-Wesley, Reading, MA) is such a separation device. It is known that the concentration per cell of a species decreases significantly during a separation of species, the original concentration having been distributed over many cells.

The fundamental principle of operation of a Feynman ratchet is based on differences in diffusivities of a number of charged species. An electrical potential is established within the device that is periodic but asymmetric in space. When the potential is on, particles are transported to and trapped in the potential wells. When the potential is off, the particles diffuse freely. Rapidly diffusing particles will be transported to an adjacent potential well during the next "on" period, while those with low diffusivity will remain approximately stationary. Thus separation occurs. Bier and Astumian (M. Bier and R.D. Astumian, 1996, *Bioelectrochem. Bioenerg.* 39:67; 1996, *Phys. Rev. Lett.* 76:4277) have presented a simple embodiment of such a device with a substantially linear array of interdigitated electrodes connected in simple fashion to a voltage source. All that is required is an inter-cell diffusion process, an intra-cell transport process, and an offset or asymmetry between the transport foci (diffusion centers) and the cell boundaries. Details of the function of the devices of the invention can be understood in this light.

The present invention provides improved versions of a Feynman ratchet. One advantage of the designs disclosed herein is that the separated species distributed over many cells can be refocused to points that are different for different species. This enhances greatly the detectability of those species by gathering what would be a weak signal distributed over many cells to a strong signal localized to a few cells in a position that depends on the species. Figures 20a and 20b (and also 22) present a graphical depiction of the refocus effect on concentration, and device design accomplishing such refocus.

This refocus capability also enhances the utility of the device for size fractionation. Such refocusing might be desirable, for example, in using the device to size-fractionate a hybridization probe (e.g. in case one wanted to see hybridization results only from the 100 to 500 base range) prior to hybridization on an integrated liquid phase size fractionation/hybridization chip.

Another advantage of the invention is that the electrode design can cause the analyte continuously to be directed toward the centerline of the separation channel (this is only known in the prior art for cases other than the use of electric potential to drive the Feynman ratchet effect--it has been described for both the optical (A. Ashkin, 1970, *Phys. Rev. Lett.* 24:156; A. Ashkin, 1986, *Optics Lett.* 11:288; L.P. Faucheux, 1995, *Phys. Rev. Lett.*

74:1504) and electric polarizability (D. Long et al., 1996, *Phys. Rev. Lett.* 76:3858; J. Rousselet et al., 1994, *Nature* 370:446) versions of the device).

Another advantage of the designs of the invention is that the material that is in contact with the electrolyte can be substantially the same all around the separation channel, and that the contact between the charged particles and the electrodes can be minimized. Certain charged particles are sensitive to chemical interactions with surfaces within the separation channel (e.g. DNA). Thus it may prove highly desirable to minimize the charged-particle-electrode interactions. The designs presented accomplish this.

A further advantage is that in one embodiment of the designs presented herein, the electrodes are not interdigitated so that for small electrode feature size fabrication of the device will not be hindered by low yield due to short-circuits between adjacent electrodes.

The invention is useful in causing motion of DNA driven by the Feynman-ratchet mechanism. The device of the invention has utility in enhancing all manner of separations and motions using Feynman ratchets. The device can be used for enhancing detectability. The device can be used for size fractionation and collection (either as a precursor to another analysis technique, such as DNA hybridization, or in and of itself). The design may be transferable to an optical Feynman ratchet design such as has been described above, optionally using holographic diffraction gratings to generate patterns of light constituting optical tweezers.

An embodiment of a Feynman ratchet device of the invention is shown in figure 18, and is referred to as the "quad-symmetric" device or electrode design. Linear interdigitated electrodes 120 are used. They are connected to at least four bond pads 130. The bond pads are attached to a voltage supply with switching capability via electrical leads or probes in a cross-wired fashion (e.g. positive in upper left and lower right, negative in upper right and lower left). Between the bond pads is a center focus gap 110. The electrode pattern can be constructed of Pt with a Ti sticking layer causing it to adhere to a substrate such as quartz or oxidized Si. Standard techniques of microfabrication can be used to achieve the desired electrode pattern in a size range from 1 to 100 micron for the electrode width.

Without loss of generality, a preferred embodiment for operation of the device will be described by way of example for the case of negatively charged DNA analyte. The device is first cleaned or treated to prevent sticking of the analyte to the device (S. Henck, U.S. Patent Application Ser. No. 60/067,387, filed December 3, 1997, which is incorporated by reference herein in its entirety). A small quantity of dilute DNA solution is applied to the surface of the device (e.g. 2 microliters at concentration 4 picomoles per microliter). Next a cover slip is used to form a fluid layer of depth less than or roughly equal to 10 microns. Sealing compound such as UV curing adhesive is applied to all edges of the cover slip. The device is then operated in usual fashion, wired as described above, in order to gather

substantially all of the DNA to the center electrodes (a bias other than zero in the periodic application of potential to the device can result in more rapid focusing of the DNA to the center focus gap 110). Finally, separation occurs when the voltage supply is switched to supply positive voltage to the upper right and lower left bond pads—the separation
5 occurring via the usual Feynman ratchet mechanism (with zero bias to the periodic potential). The separated species can be detected via standard optical techniques, such as fluorescence microscopy.

An optical pattern related to the quad-symmetric electrode design can be formed with a high power laser, beam expansion and focusing lenses, a holographic diffractive element,
10 and other optical components known in the art. Such a pattern may be used for achieving separations such as are described in A. Ashkin, 1970, *Phys. Rev. Lett.* 24:156; A. Ashkin, 1986, *Optics Lett.* 11:288; and L.P. Faucheux, 1995, *Phys. Rev. Lett.* 74:1504.

Another electrode design is illustrated in figure 19 which is similar to the quad-
15 symmetric design, having a center focus gap 110, electrodes 120, and bond pads 130 as before. In this case the electrodes contain a right angle along the diagonal of the device. This right angle alters the potential throughout the device in such a way that a charged particle located slightly off the diagonal will move toward the diagonal as it is put in motion toward the upper left corner of the device. The advantage is that DNA will remain
20 concentrated along the centerline throughout the separation process and thereby be easy to detect following separation. The design shown can be manufactured by use of a rectilinear mask design (angles require extensive mask work). One can achieve a similar effect as shown in figure 22 by allowing angles in the electrode mask patterns.

As indicated above it is desirable to be able to detect samples easily after separation,
25 and also desirable to gather samples following fractionation. The device design shown in figure 20a facilitates such detection and gathering. In the separation section 330, samples are loaded and separated as described above. With suitable application of voltages from the voltage supply and switching controller 310 via the electrical leads 320 to the distinct bond
30 pads 130 (at minimum five of them on each side of the device), the samples are caused to move further to the refocus sections 340 and 350. With suitable design parameters, e.g. for the case of two species present in the sample, a time is reached when the species are substantially separated and each lies within one of the two refocus sections. The controller
35 is then switched so that each of the refocus sections acts as a quad-symmetric device by itself. This causes the separated samples to accumulate to distinct regions in the center focus gaps 110 found at each refocus section 340, 350. The principle of operation is illustrated in figure 20b. Following loading, sealing and focusing as described above, an accumulation of each of two species can be found to the left of the device, with concentration indicated by the curves 380. After separation, the initial concentration is distributed over many cells of

the device at low concentration per cell, as shown by curves 384 and 388. Each species resides substantially within its associated refocus section 340, 350. After switching the voltage controller, the species are refocused to distinct regions at high concentration as demonstrated by curves 392 and 396. These separated and concentrated fractions of the original sample can be detected easily, or moved off the device for further use or analysis.

In the designs of the device described in detail hereinabove, there is substantial contact between the electrodes and analyte. It is believed that this contact is critical to successful use of the device. However, it is also desirable to control the surface interactions of the analyte and separation channel. With these considerations in mind, a design is presented in figure 21 which minimizes the electrode-analyte contact. A base substrate of suitable material is selected, machined or micromachined, and treated in such a way that it has suitable surface properties with respect to the analyte (e.g. quartz with RCA clean for DNA analysis—see S. Henck, U.S. Patent Application Ser. No. 60/067,387, filed December 3, 1997, which is incorporated by reference herein in its entirety). Si wafers with oxide or other coating and Pt electrodes are micromachined as has been described above. The wafers are bonded in place as shown in the perspective view. Finally, a cover of suitable material is machined to include a separation channel, treated, cleaned and bonded to seal the device. The width of the channel is slightly larger than the exposed base substrate so that the channel overlaps the edges of the wafers. The result is electrode-sample contact areas which are small. This is apparent in considering the top view and detail view shown in figure 21. Loading and operation of the device are as above. Note that in this design the electrodes are not interdigitated so that for small electrode feature size, fabrication of the device will not be hindered by low yield due to short-circuits between adjacent electrodes.

Combining the benefits described in relation to figures 18 through 20 above, a preferred embodiment of the electrode design is presented in figure 22. It is described in detail below.

An example of the preferred embodiment is shown in top view in figure 22. A short electrode segment abuts a separation section, together forming a focus section with a focus gap where samples are initially concentrated following loading and sealing of the device. A voltage supply and switching control unit is used to change the potential experienced by the sample via electrical leads. This is done in such a way that the sample is separated into its constituent species as it traverses separation section. The sample is caused to move off the end (to the right, as shown) of the separation section onto a number of refocus sections (five are shown in the figure). Ideally, the number of refocus sections substantially corresponds to the number of different species expected to be

contained in the original sample, or a number of size ranges desired for fractionation. With techniques of microfabrication this number could be as high as hundreds.

The quad-symmetric electrode design has several possible embodiments according to the details of the electrode structure near the center focus gap 110. The bond pads 130 are typically cross-wired, e.g. with positive in the upper left and lower right, as shown in figure 18. In that figure the center electrode gap shows a ground electrode on top, and a positive electrode on bottom. In alternative embodiments, a single electrode is removed from the bottom of the upper half of the device, a single electrode is removed from the top of the bottom half of the device, or both are removed. This changes the nature of the device by influencing whether the center electrode gap is bounded by closely spaced electrode pairs, or single electrodes, and by changing the charge that will be found on the center-most electrode while the analyte is being focused toward the gap 110. In a preferred embodiment, e.g. for the case of negatively charged analyte (DNA), the top electrode from the bottom half of the device is removed. During focusing of the analyte toward the gap 110, the center of the device will contain a positively charged electrode, surrounded by two single (unpaired) negatively charged electrodes. This is desirable to achieve best focusing of the negatively charged analyte to the center gap 110.

6. EXAMPLES

6.1. Separation Of Single Stranded DNA

Using the preferred model of the invention described in Sec. 5.3, the behavior of a separation device is calculated. The following device design parameters are assumed: $L = 10\mu\text{m}$, $R = 1\mu\text{m}$, length 1 cm, a potential difference of 1 V, and an aqueous separation medium. A $t_{on} = 1\text{ msec}$ and a $t_{off} = 60\text{ msec}$ are calculated as optimum for providing 2 base resolution in separating 100 base ssDNA. The total separation time is 60 min. With these design and operational parameters, the behavior of the device is calculated for a mixture of ssDNA fragments of lengths 10, 20, 30, 40, 50, 60, 70, 80, 90, and 100 (known as a standard 10-base sequencing ladder available from Research Genetics, Huntsville, Al).

Fig. 17 illustrates the predicted behavior of the device. The horizontal axis records increasing total separation time, and the vertical axis records the concentration of DNA exiting the device. The graph illustrates the predicted concentration of DNA exiting the device as a function of separation time. It is apparent that all the DNA fragments should be clearly separable.

7. SPECIFIC EMBODIMENTS, CITATION OF REFERENCES

The present invention is not to be limited in scope by the specific embodiments described herein. Indeed, various modifications of the invention in addition to those

described herein will become apparent to those skilled in the art from the foregoing description and accompanying figures. Such modifications are intended to fall within the scope of the appended claims.

Various publications are cited herein, the disclosures of which are incorporated by
 5 reference in their entireties.

8. COMPUTER PROGRAM FOR SELECTING OPTIMAL PARAMETERS

```

/*
    calculate parameters for the dna separation device
    Copyright 1996 Curagen Corporation
*/

#include    <math.h>
#include    <stdio.h>
#include    <string.h>
#include    <stdlib.h>

15  #define ABS(x) ((x)>0?(x):(-(x)))

    #define PI 3.141592653589793 /* why not? */
    #define EBREAK 1.e4 /* breakdown field for water */

    #define MINLOG -6.
20  #define MAXLOG 1.

    /* erfc by polynomial approximation */
    #define A1 0.2548296
    #define A2 -0.28449674
    #define A3 1.4214137
    #define A4 -1.453152
25  #define A5 1.0614054
    #define QP 0.3275911
    #define
        ERFC(x) (((A5*(1./(1.+QP*x))+A4)*(1./(1.+QP*x))+A3)*\
        (1./(1.+QP*x))+A2)*(1./(1.+QP*x))+A1)*(1./(1.+Qp*x))*exp(-x*x))

30  double R,L; /* R is the small spacing, L is the well spacing*/

    double alphafn(double d, double t)
    {
        double alpha,x;
        x = R sqrt(4. * d * t);
        alpha 0.5 * ERFC(x);
35  return(alpha);
    }

    main()
    {
        double n,dn; /* n and delta n */
        int nstrand; /* 1 for ssDNA, 2 for dsDNA */
  
```

```

double d,d1; /* diffusivity for length n and (n + dn) */
double logt,t,alpha,alphal,dadn,dndasq,cycles,time;
double tbest, timebest,t_on,vO;
int ntmp;
double dtmp;

5  char line[100];

    char *datafile = "data";
    FILE *fp;

    R = 1.;
    L = 10.;
10  printf("Enter R (smaller spacing) and L (larger spacing) in microns: ");
    fgets(line,sizeof(line),stdin);
    sscanf(line,"%lf %lf",&R,&L);

    R *= 1.e-4; /* convert to cm*/
    L *= 1.e-4;
15  printf("Device size: R = %lf microns, L = %lf microns\n",
    R*1.e4,L*1.e4);

    /*
        the breakdown field of water is 1e4 V/cm
        choose VO so 2VO/r = 1e4 V/cm
    */
20  vO + R * EBREAK / 2.;
    /*
        use a maximum overpotential of 2VO = 1 V to avoid electrolysis
    */
    if (vO > 0.5) { vO = 0.5;

25  printf("V_O = %lf V, generating maximum field of %lf V/cm\n",
    vO,2.*vO/R);

    printf("N = the length of the sequence\n"
    "Delta N = the resolution (1 for sequencing)\n")

    while (1) {
30  printf("Enter N and Delta N: ");
    fgets(line,sizeof(line),stdin);
    sscanf(line,"%lf %lf",&n,&dn);
    if (n < 1) { break;}
    printf("Enter 1 for ssDNA or 2 for dsDNA: "); fgets(line,sizeof(line),stdin);
    sscanf (line, "%d",&nstrand);
    if ((nstrand!=1)&&(nstrand!=2)) { break; }
35  fp = fopen(datafile,"W");

    /*      determine the diffusion constants for n and n+1 units are cm-2/sec
    t-on is the relaxation time for a 200 V potential
    */
    if (nstrand == 1) {
        t_on  (L-R)*(L-R)*2.24e-4/(R*pow((double) n,0.41));

```

```

    t_on *= 1.e4; /* convert from cm to microns */
  }
  else {
    t_on (L-R)*(L-R)*1.12e-4/R;
    t_on 1.e4; /* convert from cm to microns */
  }
5  printf("t-on + %1f s\n",t_on);
  if (nstrand == 1) {
    d = 1.14e-6 * pow(n, -0.59);
    d1 = 1.14e-6 * pow(n+dn, -0.59);
  }
  else {
    d = 1.14e-6 / n;
10 d1 = 1.14e-6 (n+dn);
  }

  tbest = 0.; timebest = 1.e100;
  for (logt = MINLOG; logt <= MAXLOG; logt += 0.001) {
    t = pow(10.,logt);
    alpha = alphafn(d,t);
15  alpha1 = alphafn(d1,t);
    dadn = alpha1 - alpha;
    if (ABS(dadn) < 1.e-6) {continue;}
    dndasq = 1. / (dadn * dadn);
    cycles = alpha * (1. - alpha) * dndasq;
    time = cycles * (t + t_on);
    fprintf(fp, "%1f %1f %1f %1f %1f %15.101f\n",
20  t,time/60.,cycles,alpha,dndasq,dadn);
    if (time < timebest) { tbest = t; timebest = time;}
    if (time > 10.*timebest) {break;}
  }
  fclose(fp);

  t = tbest;
25  alpha = alphafn(d,t);
  alpha1 = alphafn(d1,t);
  dadn = alpha1 - alpha;
  dndasq = 1. / (dadn * dadn);
  cycles = alpha * (1. - alpha) * dndasq;
  time = cycles * (t+t_on);
  printf(" N %d Delta %d nstrand %d R (um) %1f L (um) %1f\n",
30  (int)n,(int)dn,nstrand,1.e4*R,1.e4*L);
  printf(" t_on %g vO %g\n",t_on,vO);
  printf("\nN = %d +/- %d\n"
    "time (min) %1f\n"
    "length (cm) %1f\n"
    "t_off(sec) %1f\n"
    "N_cyc %f\n"
35  "alpha %1f\n\n",
    (int)n,(int)dn,
    time/60.,alpha*cycles*L,t,cycles,alpha);

  printf("%1f %1f\n",1.e4*L,alpha*cycles*L);
  printf("%1f %1f\n",t+t_on,time/60.);
  for (ntmp = 10; ntmp <= 100; ntmp += 10) {

```

```
dtmp = 1.14e-6 * pow(ntmp, -0.59);  
printf("%d %lf\n",ntmp,alphafn(dtmp,t));  
}
```

```
printf("End of program.\n");
```

```
5 }
```

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WHAT IS CLAIMED IS:

1. A method for focusing a packet of charged particles in a charged medium
5 comprising exposing the particles in the medium to an electric potential having spatial and temporal variations comprising a plurality of potential wells, each having a well minimum, said temporal variations comprising repeated variations between at least two conditions, an “on” condition in which the charged particles are attracted to the minima of the potential wells and an “off” condition in which the particles can diffuse in the medium.
10
2. A method for moving a packet of charged particles in a medium along a direction of motion, comprising:
 - (a) focusing the particles in the medium, and
 - 15 (b) exposing the focused particles in the medium to an electric potential having spatial and temporal variations comprising a plurality of potential wells, each having a well minimum, dispersed along the direction of motion and said temporal variations comprising repeated variations between at least two conditions, an “on” condition in which
20 the charged particles are attracted to the minima of the potential wells and an “off” condition in which the particles can diffuse in the medium.
3. A method for separating charged particles in a separation medium along a
25 separation direction comprising:
 - (a) focusing the particles in the medium;
 - (b) dividing the particles into a plurality of species; and
 - (c) refocusing each of said species.
30
4. The method according to claim 3, wherein the step of dividing the particles into a plurality of species is accomplished by exposing the focused particles in the medium to an electric potential having spatial and temporal variations disposed to effect division, said spatial variations comprising a plurality of potential wells, each having a well
35 minimum, disposed along the direction of separation and said temporal variations comprising repeated variations between at least two conditions, an “on” condition in which the charged particles are attracted to the minima of the potential wells and an “off” condition in which the particles can diffuse in the separation medium.

5. The method according to claim 4 wherein the time variation of the electric potential has a frequency that is varied during particle separation.

6. The method according to claim 3, wherein the step of focusing the particles in
5 the medium is accomplished by exposing the particles in the medium to an electric potential having spatial and temporal variations disposed to effect focusing, said spatial variations comprising a plurality of potential wells, each having a well minimum, disposed along the direction of separation and said temporal variations comprising repeated variations between
10 at least two conditions, an "on" condition in which the charged particles are attracted to the minima of the potential wells and an "off" condition in which the particles can diffuse in the separation medium.

7. The method according to claim 3, wherein step (c) is accomplished by
15 refocusing each of said species on distinctly addressed pads.

8. A device for separating charged particles into a plurality of species comprising:

- 20 (a) a means for focusing the particles;
- (b) one or more separation lanes capable of containing a separation medium; and
- (c) a plurality of separation electrodes situated so as to be capable of
25 generating an electric potential in said one or more separation lanes, said electric potential having a stationary spatial pattern with a time-varying amplitude.

9. The device according to claim 8, wherein said plurality of separation electrodes comprises a first sub-plurality and a second sub-plurality of separation electrodes,
30 wherein electrodes of each sub-plurality are substantially periodically spaced apart by the same distance, and wherein the first and second sub-pluralities of separation electrodes are interdigitated.

10. The device according to claim 9, wherein said plurality of separation
35 electrodes is connected to a number of bonding pads that is at least four, said bonding pads being attached to a voltage supply with switching capability in a cross-wired fashion.

11. The device according to claim 10, wherein said plurality of separation electrodes contains a right angle along a diagonal of the device.

12. The device according to claim 9 or 10, further comprising a plurality of refocus sections, each of said refocus sections comprising one or more refocusing gaps and a plurality of refocusing electrodes situated so as to be capable of generating an electric potential in said one or more refocusing gaps, said electric potential having a stationary
5 spatial pattern with a time-varying amplitude.

13. The device according to claim 12, wherein the number of said refocus sections corresponds substantially to the number of said species.

10 14. The device according to claim 12, wherein said plurality of refocusing electrodes contains a right angle along a diagonal of the device.

15 15. The device according to claim 8, wherein said separation electrodes are configured so as to minimize contact between said separation electrodes and said separation medium.

16. The device according to claim 8 wherein said electrodes are platinum and are bonded to a substrate by means of a titanium adhesion layer.
20

17. The device according to claim 8, wherein said separation lane is bounded by a first wafer and a second wafer, said first wafer having a planar first major surface and a planar second major surface, said second wafer having a planar first major surface in the same plane as the first major surface of said first wafer and having a planar second major
25 surface in the same plane as the second major surface of said first wafer, said first major surface of said first wafer and said first major surface of said second wafer bonded to a substrate, and said second major surface of said first wafer and said second major surface of said second wafer bonded to a cover so as to seal the device.

30 18. The device according to claim 8, wherein a single electrode adjacent to one or more separation lanes is not present in the device.

19. The device according to claim 8, wherein both electrodes adjacent to one or
35 more separation lanes are not present in the device.

20. A device for focusing charged particles comprising:
one or more focusing gaps capable of containing a focusing medium; and

a plurality of focusing electrodes situated so as to be capable of generating an electric potential in said one or more focusing gaps, said electric potential having a stationary spatial pattern with a time-varying amplitude.

5 21. The device according to claim 20, wherein said plurality of focusing electrodes is further configured to cause continuous concentration of the charged particles to a center line.

10 22. The device according to claim 20, wherein said plurality of focusing electrodes is further configured so as to minimize contact between said focusing electrodes and said focusing medium.

15 23. The device according to claim 20, wherein said plurality of focusing electrodes comprises a first sub-plurality and a second sub-plurality of focusing electrodes, wherein electrodes of each sub-plurality are substantially periodically spaced apart by the same distance, and wherein the first and second sub-pluralities of focusing electrodes are interdigitated.

20 24. The device according to claim 23, wherein said plurality of focusing electrodes is connected to at least four bonding pads, said bonding pads configured so that adjacent bonding pads have opposite voltage polarity.

25 25. The device according to claim 24, wherein said plurality of focusing electrodes contains a right angle along a diagonal of the device.

30

35

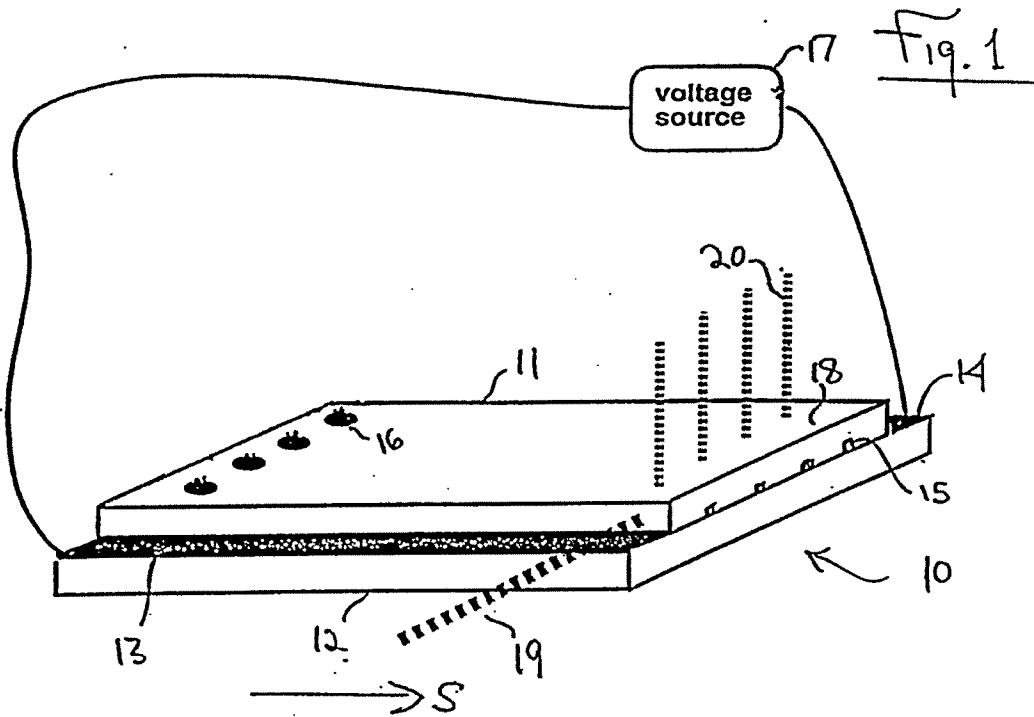


Fig. 2

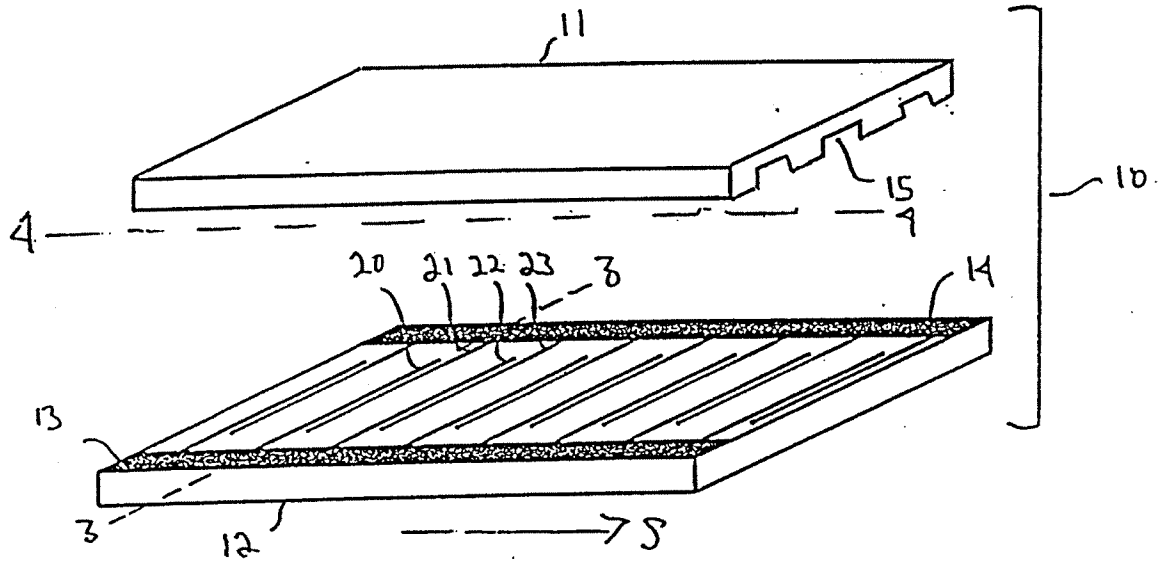
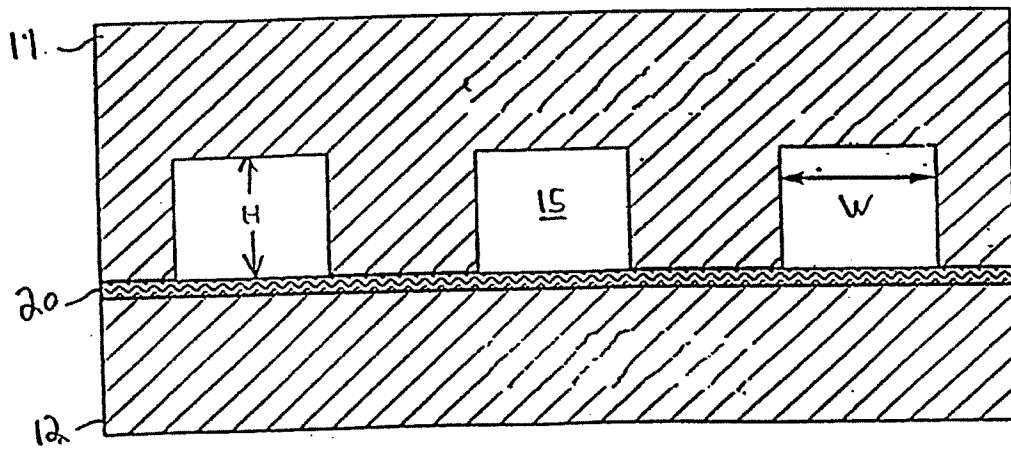


Fig. 3



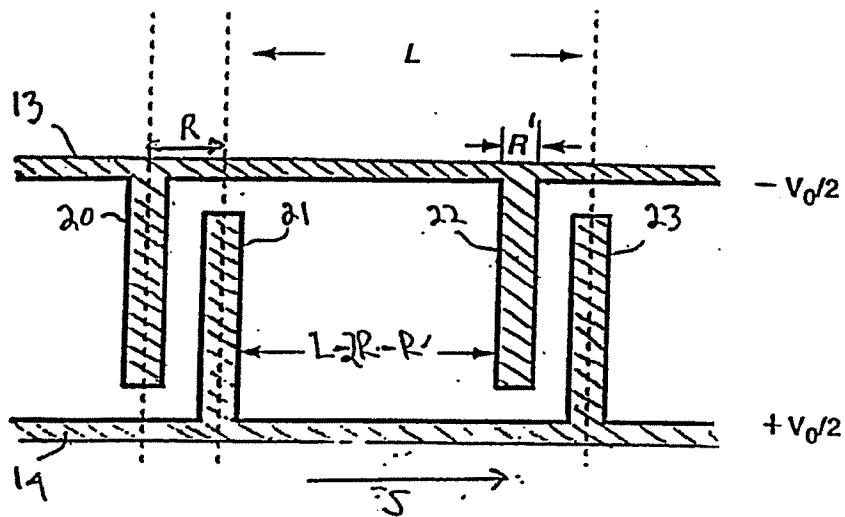


Fig. 4A

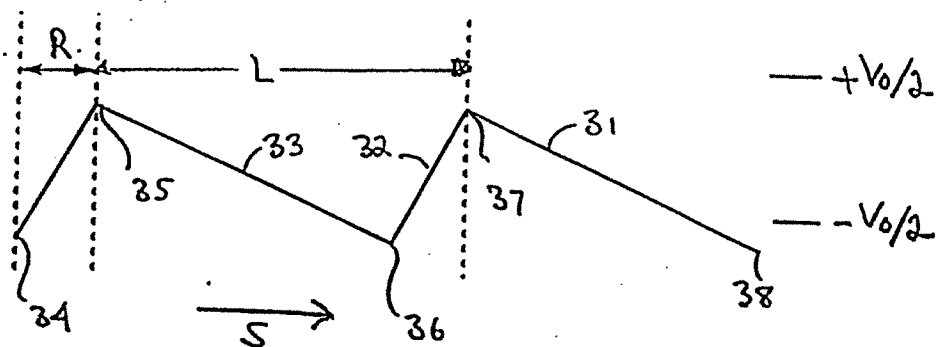
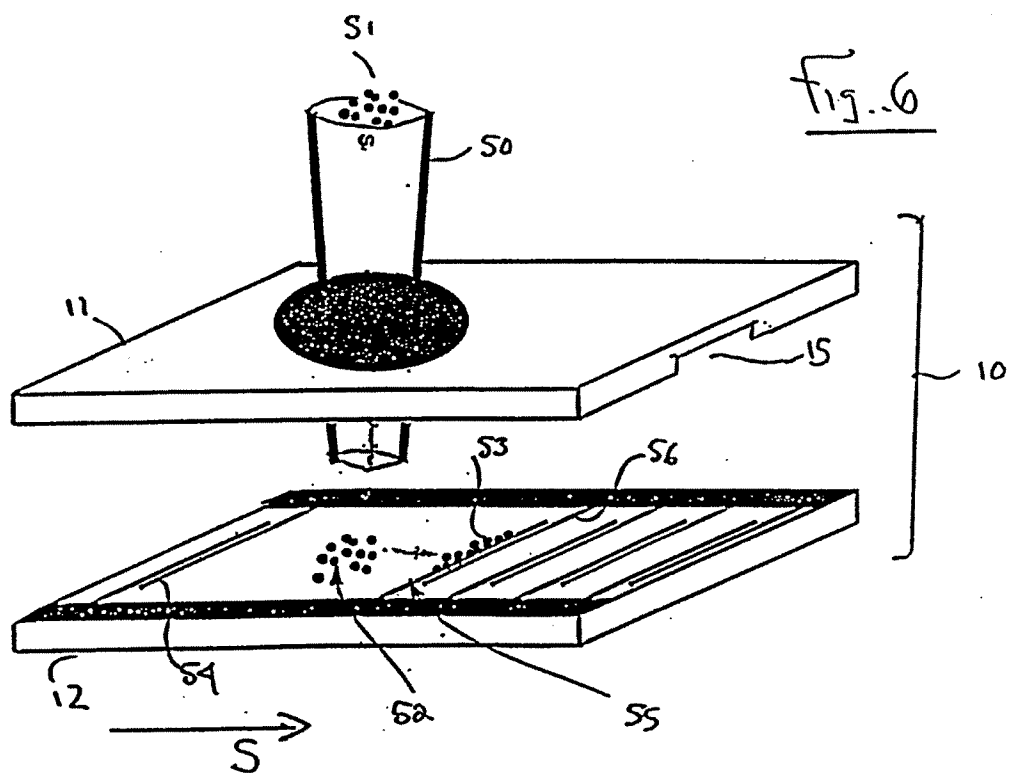
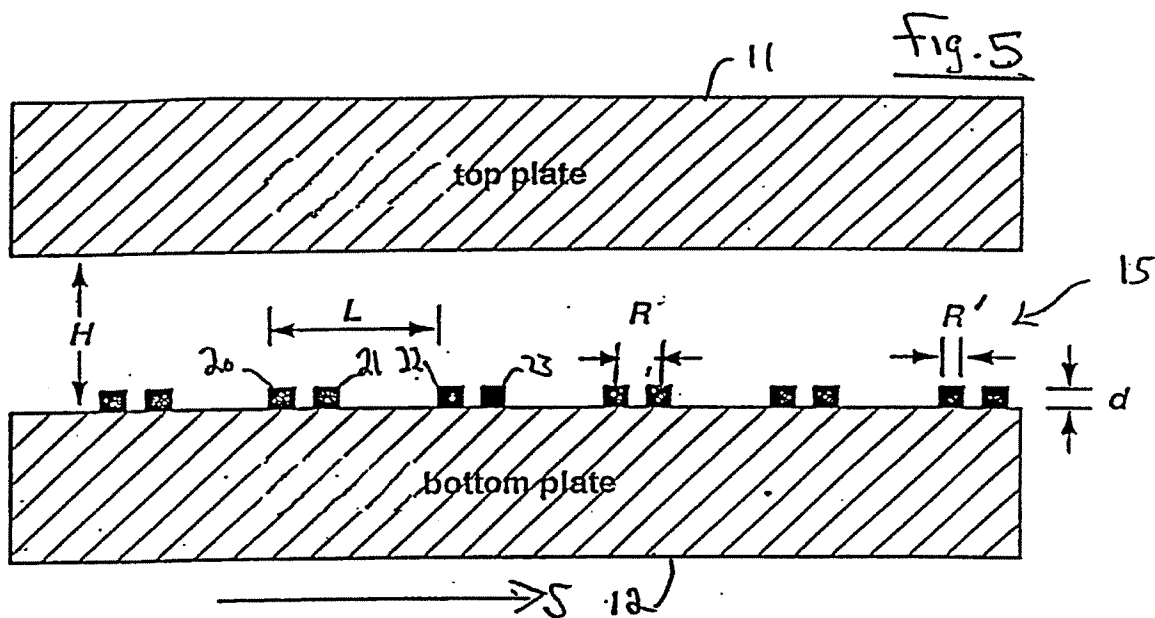
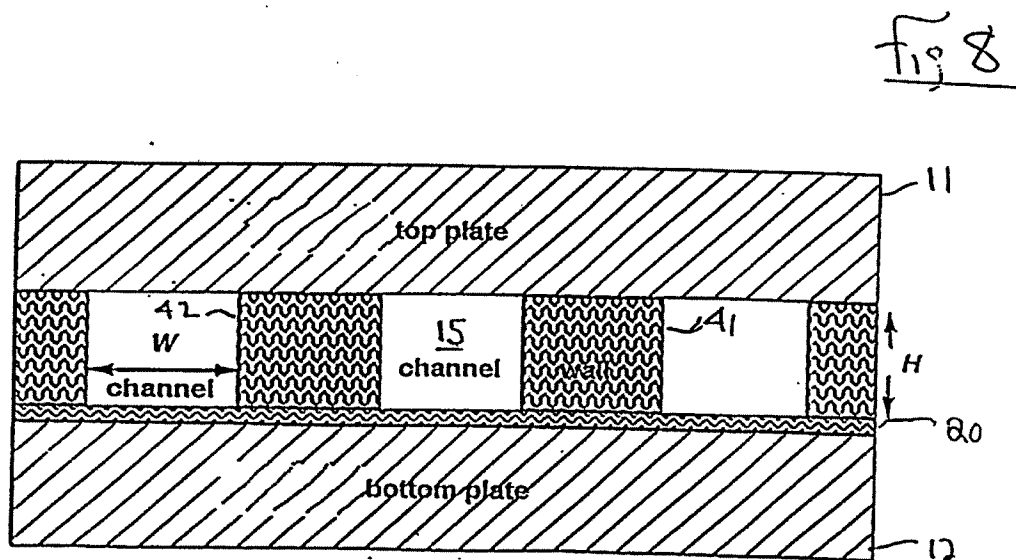
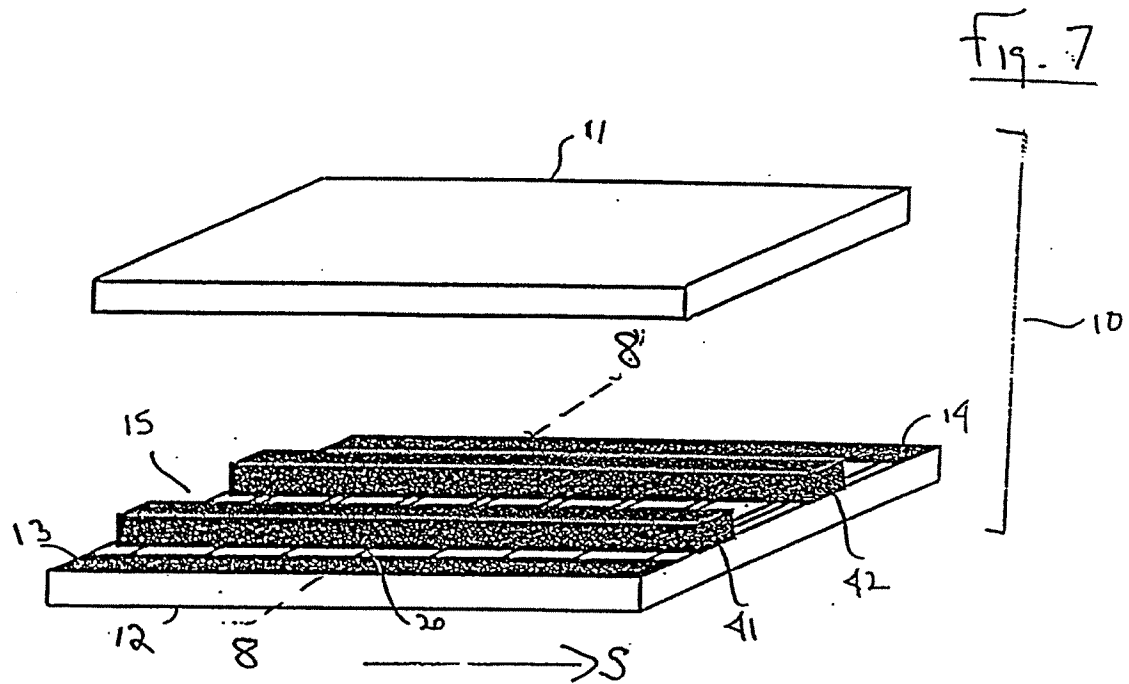
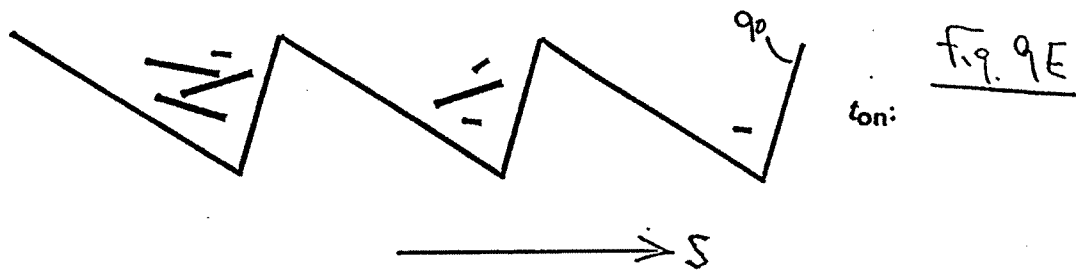
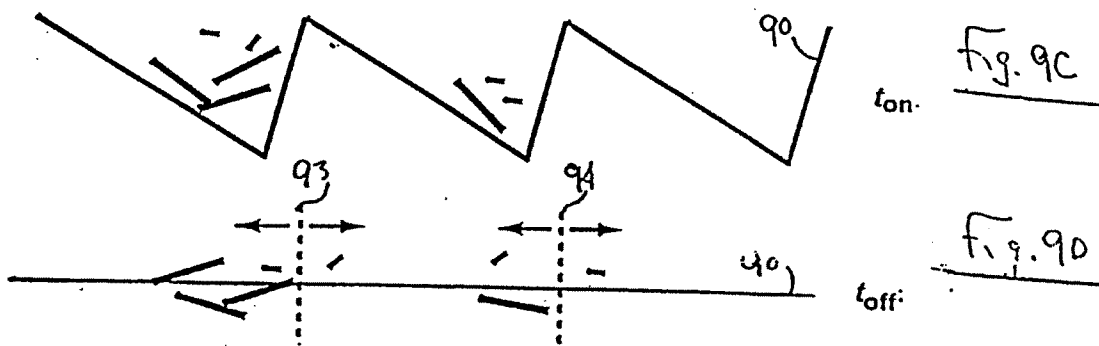
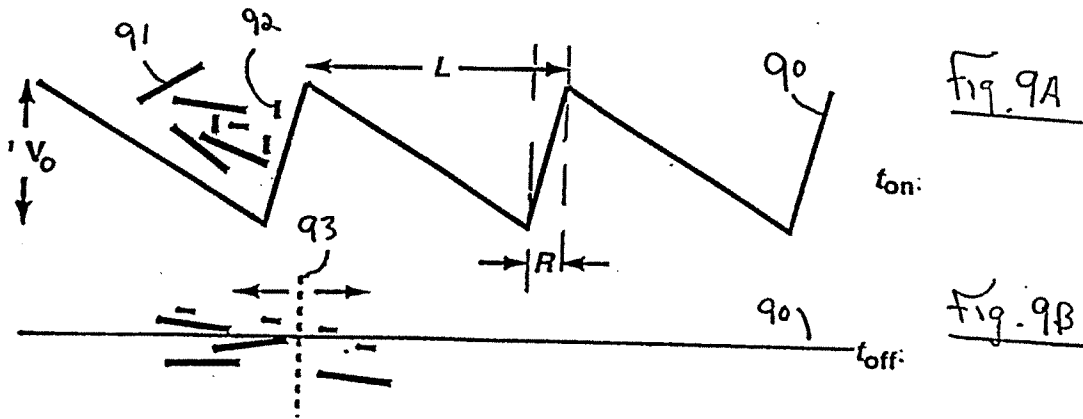


Fig. 4B







7/19

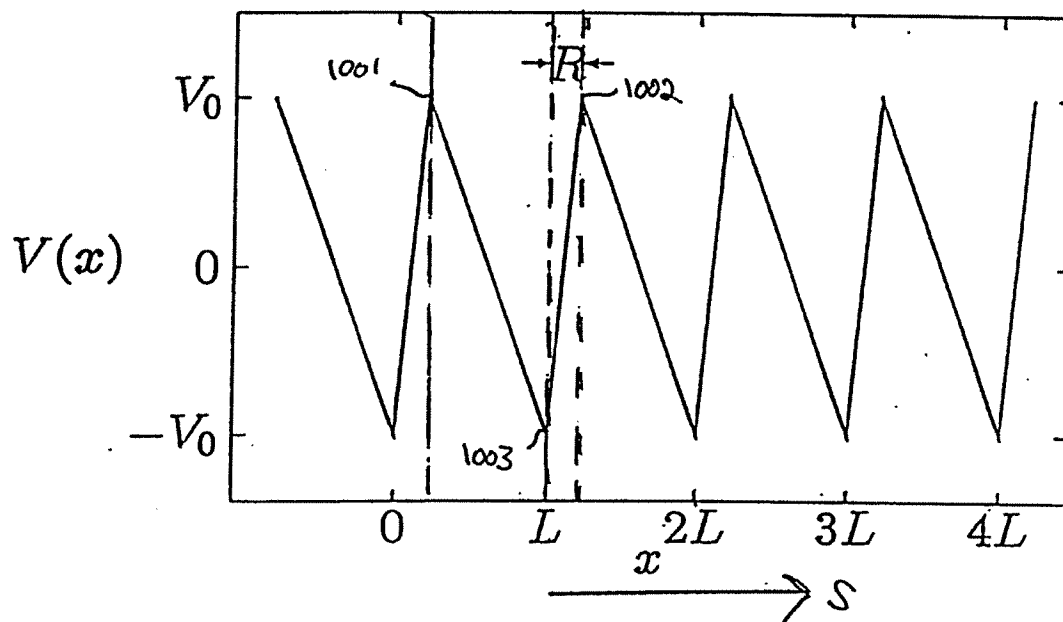
fig. 10

Fig. 11A

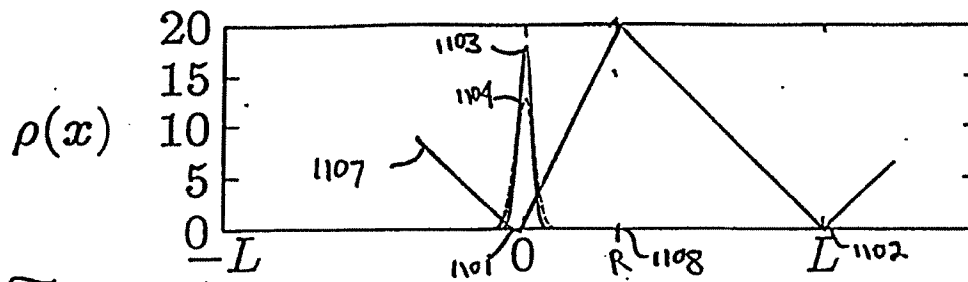


Fig. 11B

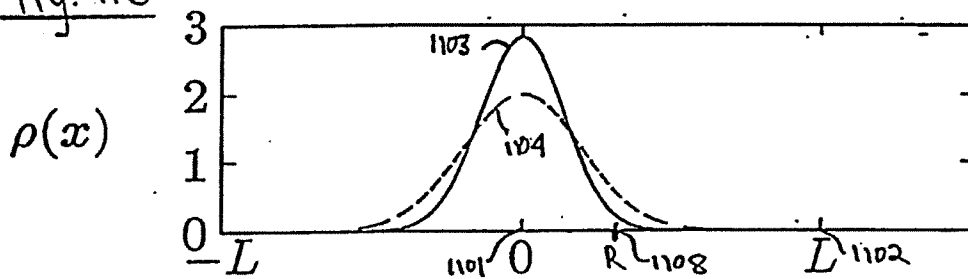


Fig. 11C

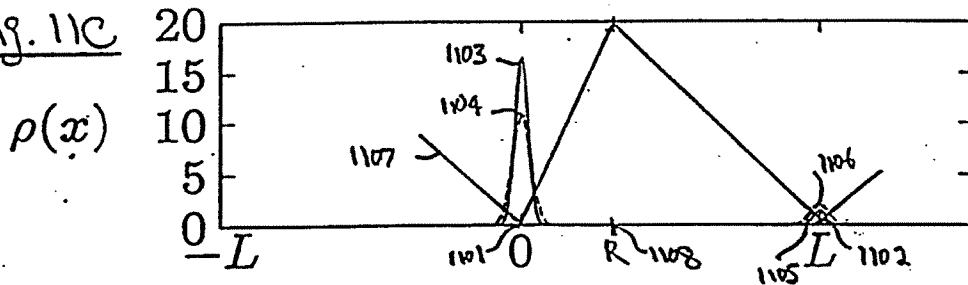
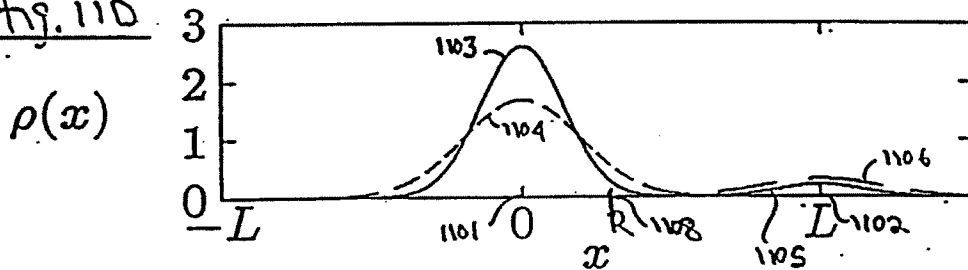


Fig. 11D



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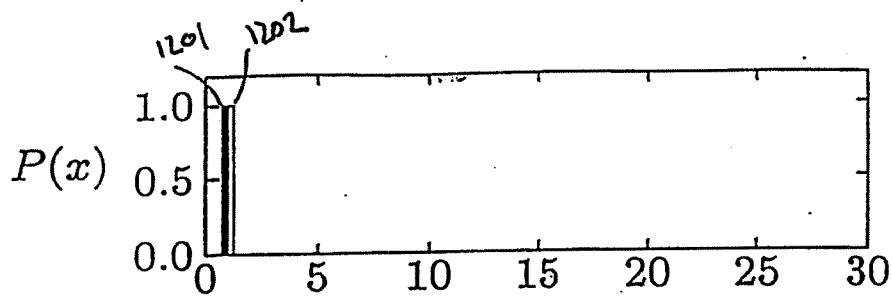


Fig 12A

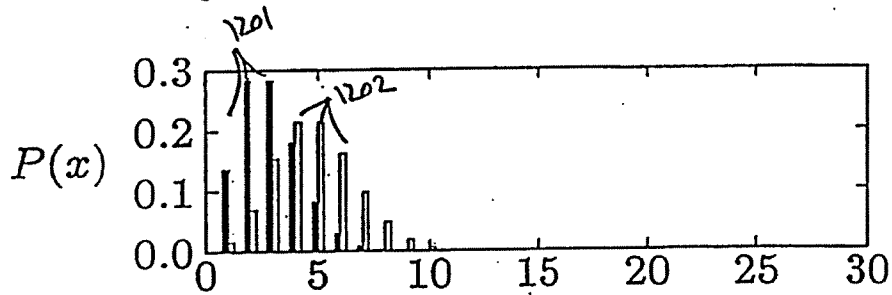


Fig 12B

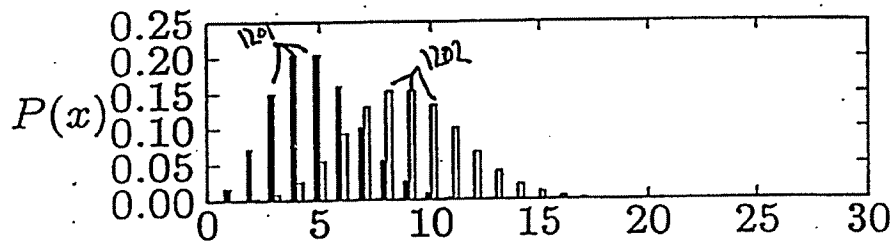


Fig 12C

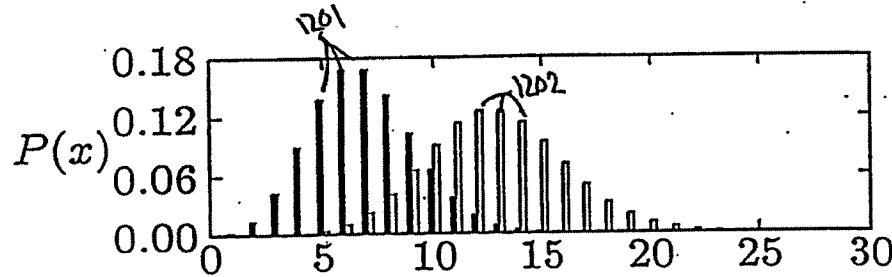


Fig 12D

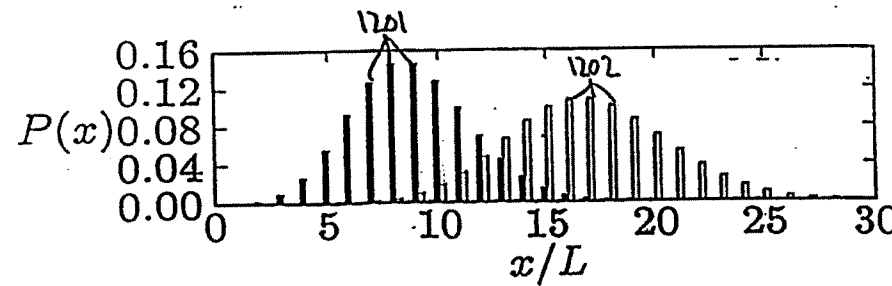


Fig 12E

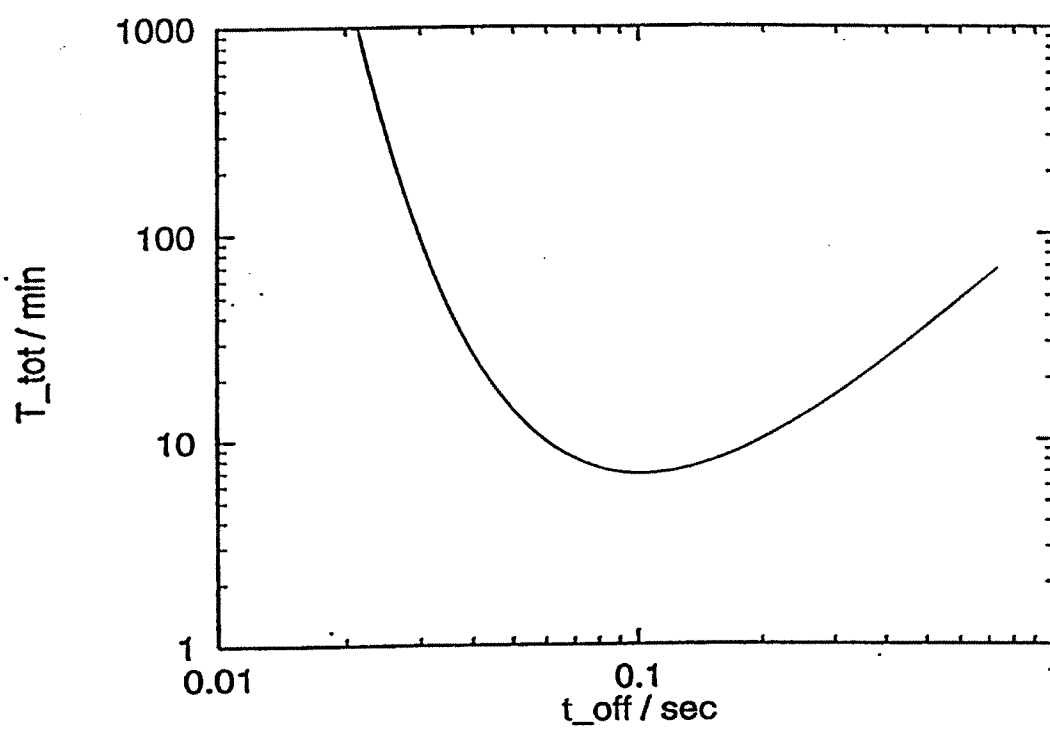
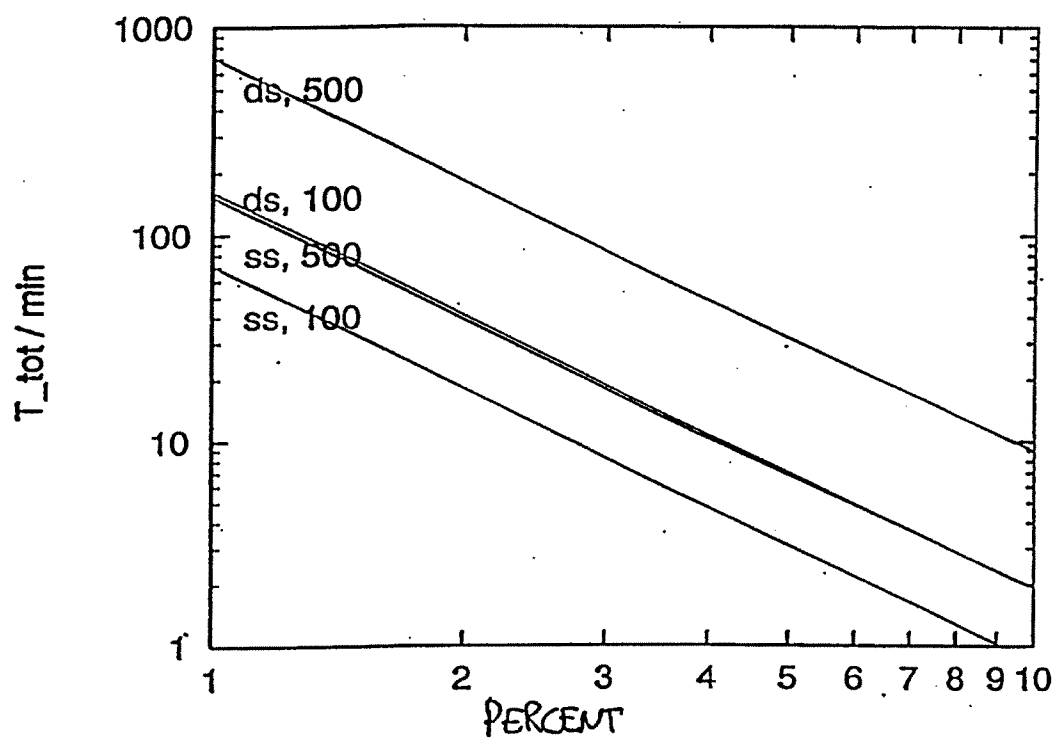
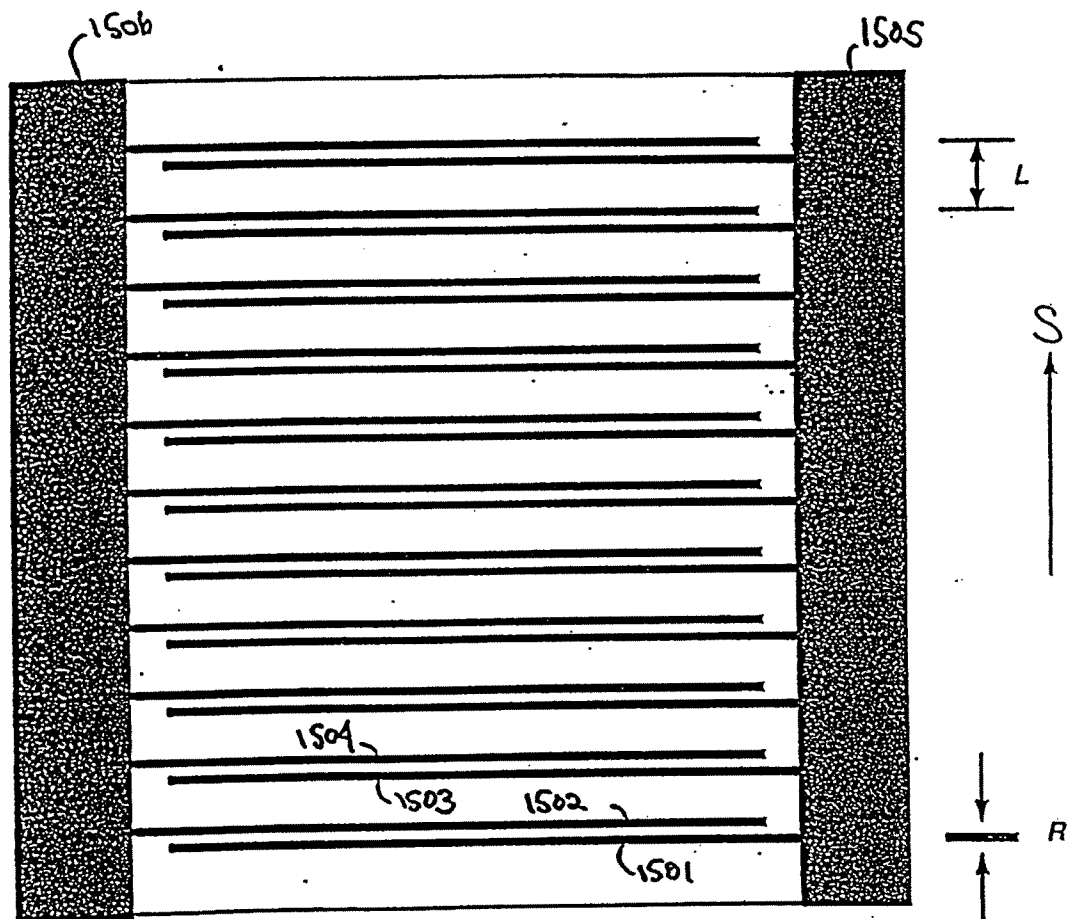
Fig. 13

Fig. 14

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Fig. 15

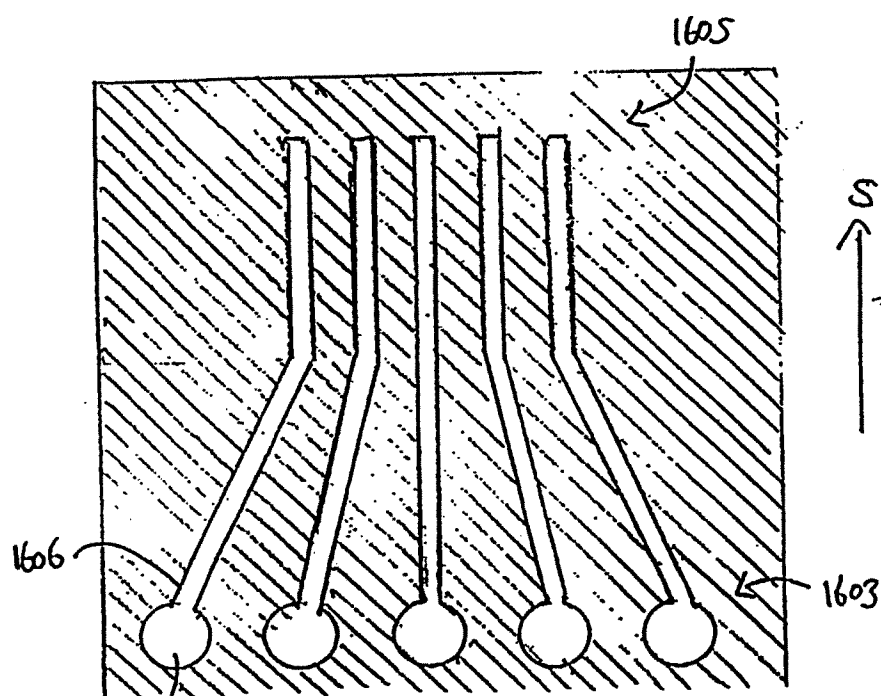
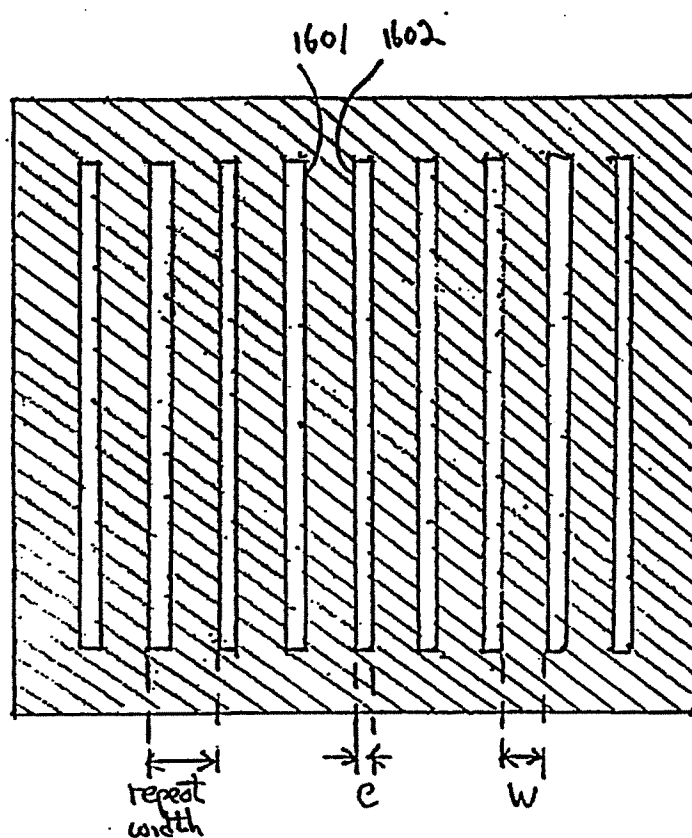
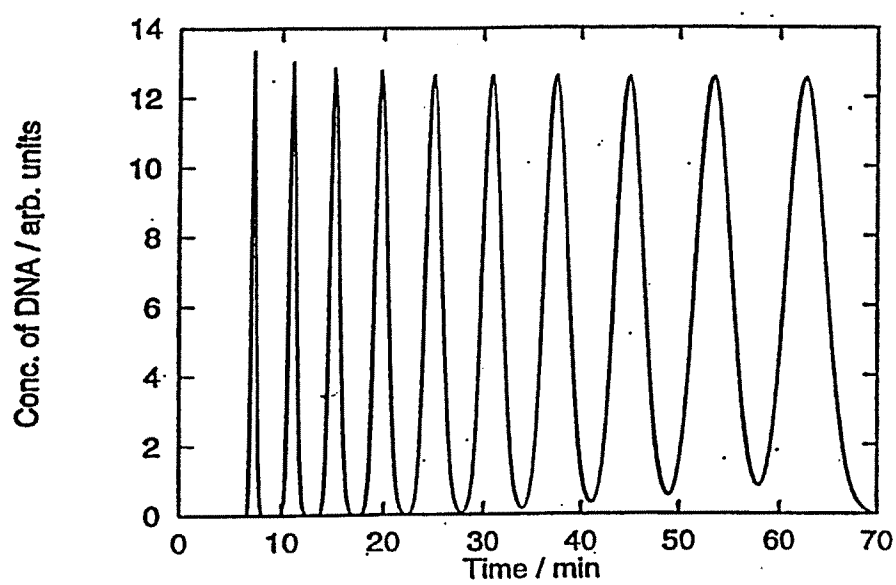


Fig. 17

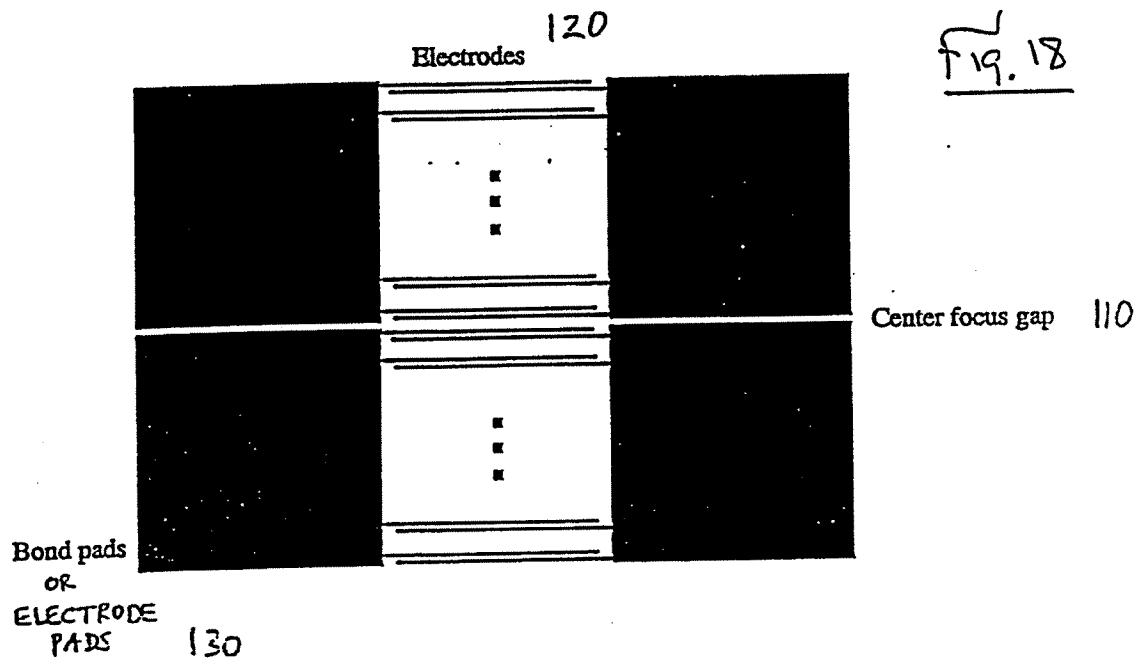


FIG. 19

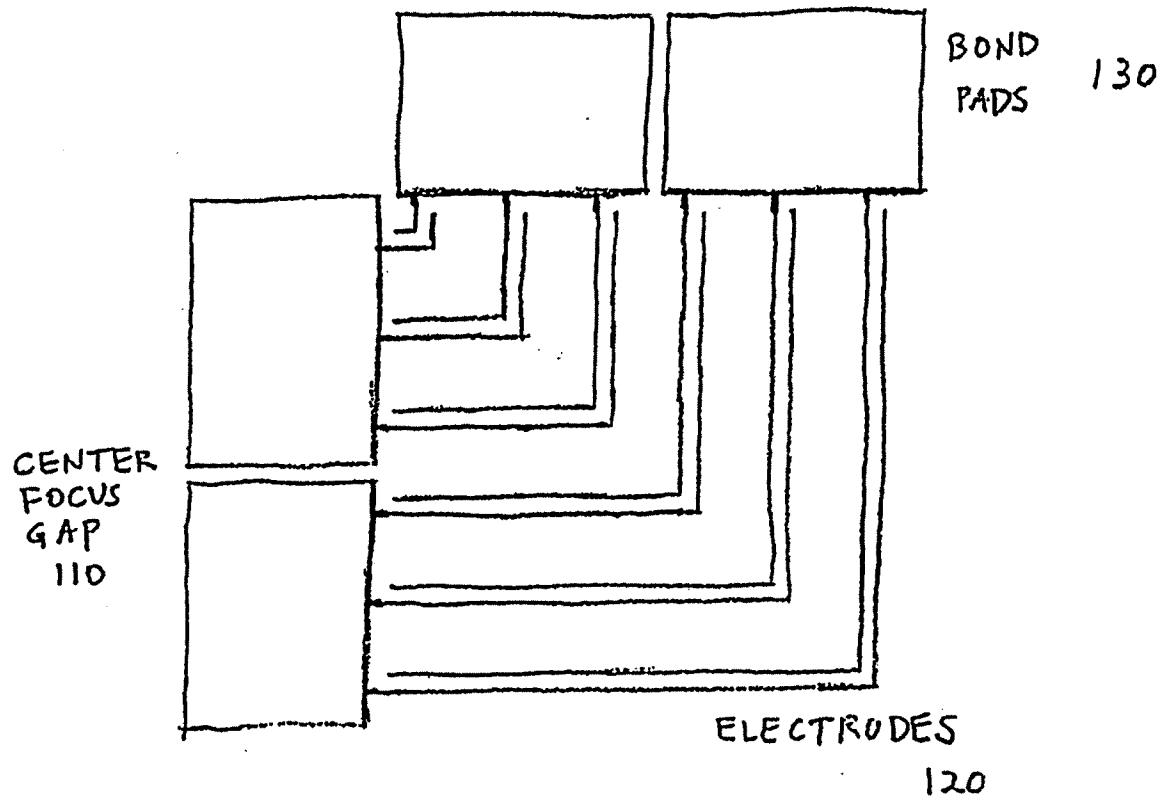


Fig. 20A

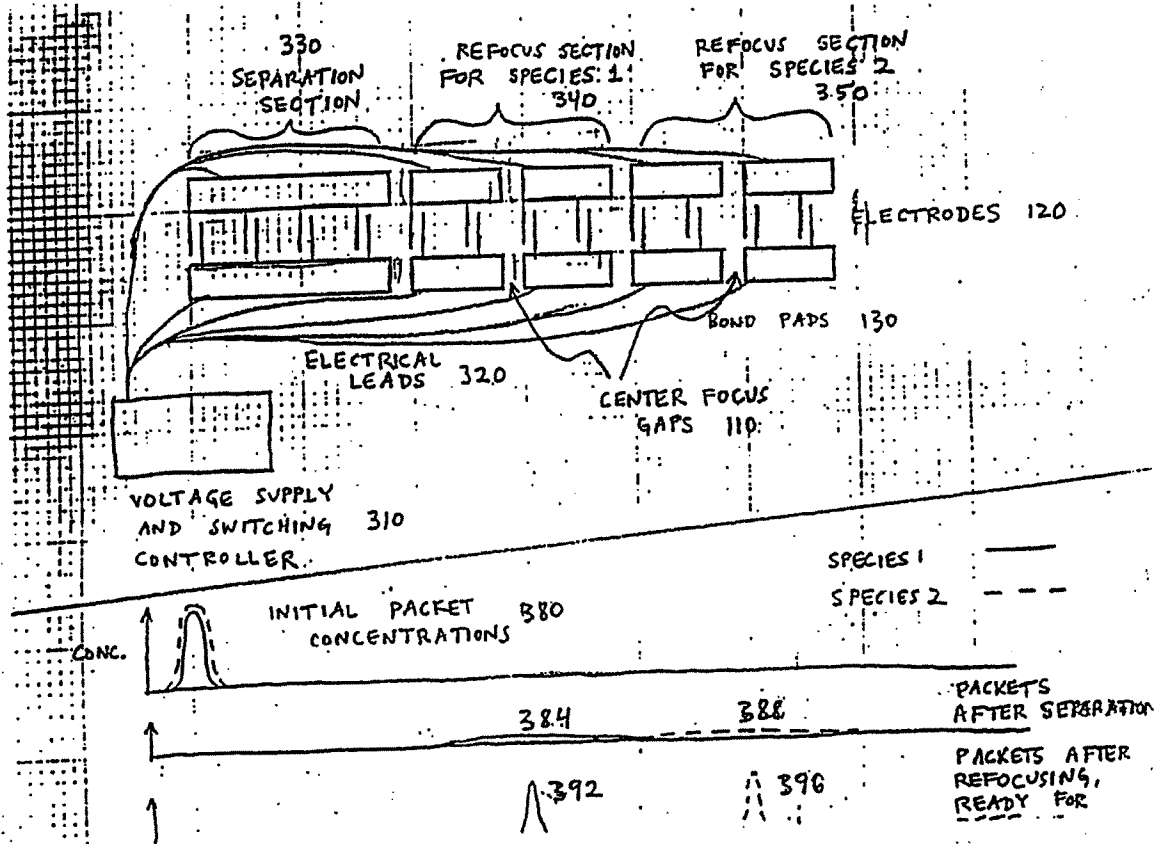


Fig. 20B

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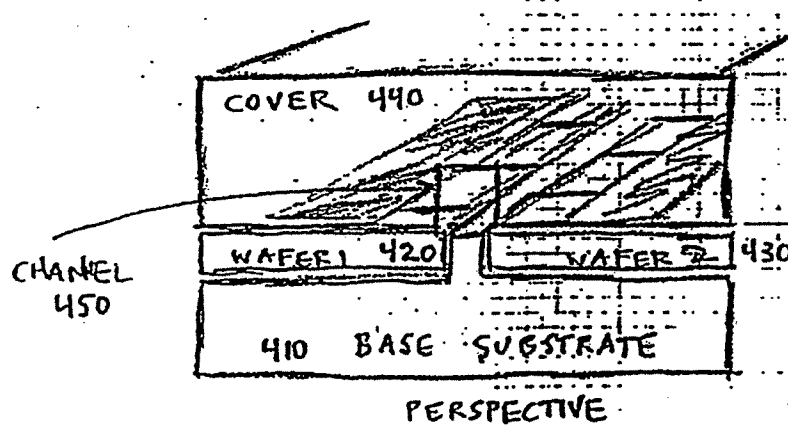
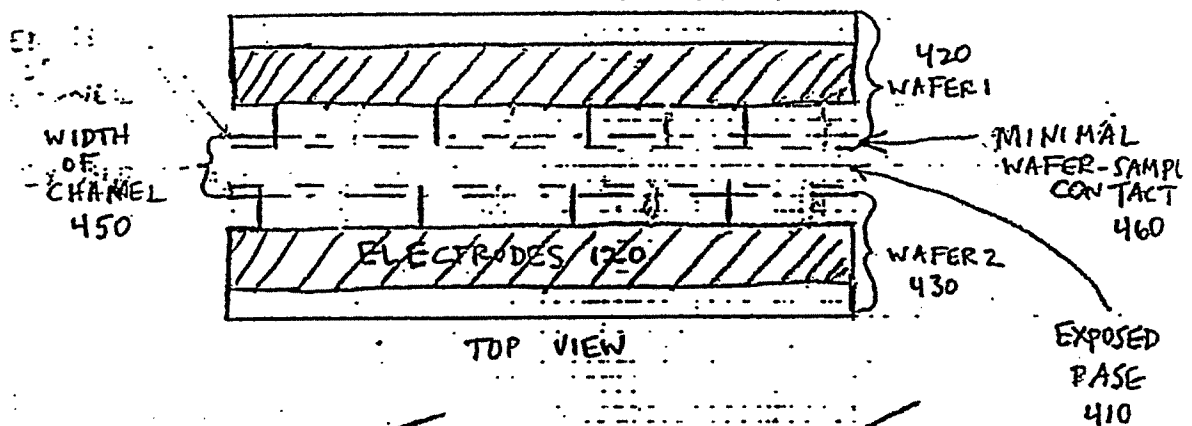
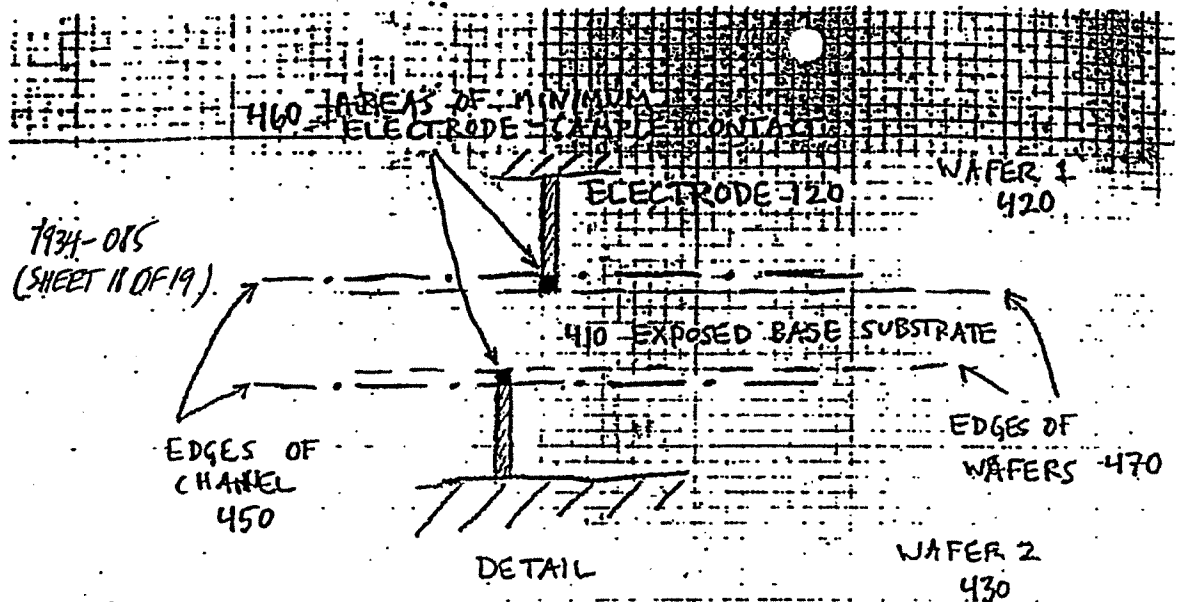


Fig. 21

INTERNATIONAL SEARCH REPORT

 International application No.
PCT/US99/29195
A. CLASSIFICATION OF SUBJECT MATTER

IPC(7) :G01N 27/26; G01N 27/447

US CL :204/450, 600

According to International Patent Classification (IPC) or to both national classification and IPC

B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)

U.S. : 204/450, 451, 547, 600, 601, 601, 603, 660,663

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

Electronic data base consulted during the international search (name of data base and, where practicable, search terms used)

C. DOCUMENTS CONSIDERED TO BE RELEVANT

Category*	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
X	US 5,569,367 A (BETTS et al) 29 October 1996, fig. 2 and col. 5, line 49- col.6, line19	8,9,11,15, 18-23,25
-----		-----
Y		16,17
X	US 5,593,565 A (AJDARI et al) 14 January 1997, see entire document	1
X	ROUSSELET, JULLETTE et al Directional Motion of Brownian Particles Induced by a Periodic Assymmetric Potential. Nature. 11 August 1994. Vol.370. pages 446-488	1
A	US 6,024,854 A (GILCHRIST) 15 February 2000	2-25
A	US 5,833,826 A (NORDMAN) 10 November 1998	3



Further documents are listed in the continuation of Box C.



See patent family annex.

* Special categories of cited documents:	"T" later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention
"A" document defining the general state of the art which is not considered to be of particular relevance	"X" document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to involve an inventive step when the document is taken alone
"E" earlier document published on or after the international filing date	"Y" document of particular relevance; the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art
"L" document which may throw doubts on priority claim(s) or which is cited to establish the publication date of another citation or other special reason (as specified)	"&" document member of the same patent family
"O" document referring to an oral disclosure, use, exhibition or other means	
"P" document published prior to the international filing date but later than the priority date claimed	

Date of the actual completion of the international search

18 FEBRUARY 2000

Date of mailing of the international search report

07 MAR 2000

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